

Diagnostics and Therapeutics for Diseases Associated with human phosphodiesterase 11A (PDE11A)

5 **Technical field of the invention**

The present invention is in the field of molecular biology, more particularly, the present invention relates to nucleic acid sequences and amino acid sequences of a human PDE11A and its regulation for the treatment of disorders of the peripheral and
10 central nervous system, cardiovascular diseases, cancer, liver disease and genitourinary diseases in mammals.

Background of the invention

15 PDE11A is a member of the enzyme family of phosphodiesterases (PDEs) [Fawcett L et al., (2000)] , WO200040733, WO200166716, [Yuasa K et al. (2000)]. PDEs catalyze the hydrolyzation of 3', 5' cyclic nucleotides. That results in the formation of the respective nucleoside 5' monophosphates. The cyclic nucleotides cAMP and cGMP serve as crucial second messengers in a number of cellular signaling
20 pathways. The PDEs as well as the guanylyl and adenylyl cyclases, which synthesize the cyclic nucleotides, are important cellular components to regulate the concentration of cyclic nucleotides and, thus, to regulate the signal transduction pathways. Because of their central role in regulating second messenger levels PDEs have been considered chemotherapeutic targets and have been worked on
25 extensively.

Several families of PDEs have been identified. The nomenclature system includes first a number that indicates the PDE family. To date, eleven families (PDE 1-11) are known which are classified by: (i) primary structure; (ii) substrate preference; (iii)
30 response to different modulators; (iv) sensitivity to specific inhibitors; and (v) modes of regulation [Loughney and Ferguson, (1996)]. The number indicating the family is

followed by a capital letter, indicating a distinct gene, and the capital letter followed by a second number, indicating a specific splice variant or a specific transcript that utilizes a unique transcription initiation site.

5 PDEs show of the following structural features:

All mammalian PDEs identified to date possess a highly conserved region of 270-300 amino acids in the carboxy terminal half of the protein [Charbonneau, et al. (1986)]. Here, the catalytic site for cAMP and/or cGMP hydrolysis and two putative
10 zinc binding sites as well as family specific determinants are located [Beavo, (1995); Francis, et al. (1994)]. The amino terminal regions of the various PDEs are highly variable and include other family specific determinants and diverse regulatory motifs such as: (i) calmodulin binding sites (PDE1); (ii) non-catalytic cyclic GMP binding sites (PDE2, PDE5, PDE6); (iii) membrane targeting sites (PDE4); (iv) hydrophobic
15 membrane association sites (PDE3); and (v) phosphorylation sites for either the calmodulin-dependent kinase II (PDE1), the cAMP-dependent kinase (PDE1, PDE3, PDE4), or the cGMP dependent kinase (PDE5) [Beavo, (1995); Manganiello, et al. (1995); Conti, et al. (1995)].

20 Members of the PDE1 family are calcium-calmodulin dependent. The group is composed of at least three genes with several splicing variants [Kakkar, R. et al. (1999)]; PDE1A and PDE1B preferentially hydrolyze cGMP while PDE1C is dualspecific, it exhibits a high affinity for both cAMP and cGMP. In vitro experiments show regulation of some PDE1 species by phosphoprylation, which
25 decreases the affinity of the enzyme for calmodulin [Kakkar, (1999)]. PDE1s have been shown to be expressed in lung, heart and brain.

The PDE2 family is characterized as being specifically stimulated by cGMP [Loughney and Ferguson, supra]. PDE2 species have been found in cerebellum,
30 neocortex, heart, kidney, lung, pulmonary artery, and skeletal muscle [Sadhu, K. et

al. (1999)]. Only one gene has been identified, PDE2A. The respective PDE2A protein is specifically inhibited by erythro-9-(2-hydroxy-3-nonyl)adenine (EHNA).

Two genes have been identified in the PDE3 family, PDE3A and PDE3B, both having high affinity for both cAMP and cGMP, although the V_{\max} for cGMP hydrolysis is low enough that cGMP functions as a competitive inhibitor for cAMP hydrolysis. Enzymes in the PDE3 family are specifically inhibited by cGMP. PDE3 enzymes are specifically inhibited by milrinone and enoximone [Loughney and Ferguson, supra].

PDE4s are specific for cAMP hydrolysis. The family is comprised of four genes, PDE4A, PDE4B, PDE4C, and PDE4D. The genes give rise to multiple splice variants and are expressed in airway smooth muscle, the vascular endothelium, and all inflammatory cells. The enzymes can be activated by cAMP-dependent phosphorylation. Members of this family are specifically inhibited by the antidepressant drug rolipram.

PDE5 is highly selective for cGMP [Turko, I.V. et al. (1998)]. Members of PDE5 family bind cGMP at non-catalytic sites [McAllister-Lucas, L.M. (1995)]. CGMP binding at non-catalytic sides has been suggested to be important for phosphorylation by cGMP-dependent protein kinase. PDE5 is highly expressed in vascular smooth muscle, platelets, lung, and kidney. Only one gene, PDE5A, has been identified.

PDE6s, the photoreceptor enzymes specifically hydrolyze cGMP [Loughney and Ferguson, supra]. PDE6s possess 2 regulatory high affinity cGMP binding sides. Genes include PDE6A and PDE6B (the protein products of which dimerize and bind two copies of a smaller γ inhibitory subunit to form rod PDE), in addition to PDE6C which associates with three smaller proteins to form cone PDE.

The PDE7 family effects cAMP hydrolysis but, in contrast to the PDE4 family, is not inhibited by rolipram [Loughney and Ferguson, supra]. Only one gene, PDE7A, has

been identified. PDE7A gives rise to multiple splice variants. PDE7 mRNA can be found in several tissues but PDE7 protein expression appears to be restricted [Han, P. et al. (1997); Perry, M.J. and G.A. Higgs (1998)]. Not much is known about the physiological function of PDE7.

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The PDE8 family is closely related to the the PDE4 family. PDE8s have been shown to hydrolyze cAMP and are insensitive to inhibitors specific for PDEs 1-5. PDE8s are found in thyroid gland, testis, eye, liver, skeletal muscle, heart, kidney, ovary, and brain.

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The PDE9 family preferentially hydrolyzes cGMP and is not sensitive to inhibition by rolipram, a PDE4-specific inhibitor, or isobutyl methyl xanthine (IBMX), a non-specific PDE inhibitor. PDE9 expression has been demonstrated in kidney, liver, lung, brain, spleen, and small intestine. Depending on nomenclature used, PDE9 is also referred to as PDE8, but is distinct from PDE8 mentioned above.

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PDE10 family members hydrolyze both cAMP and cGMP. PDE10s show expression in brain, thyroid and testis. [Soderling, S.H. et al. (1999); Fujishige, K. et al. (1999); Loughney, K. et al (1999)]

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Members of the recently identified PDE11 family are also dualspecific. Interestingly, PDE11 splice variants exhibit different regulatory sequences in the N-terminal region. This suggests the possibility of differential regulation of PDE11s [Hetman JM, Robas N, Baxendale R, Fidock M, Phillips SC, Soderling SH, Beavo JA (2000)].

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Increased PDE activity and decreased levels of cyclic nucleotides have been shown to be associated with many diseases. Furthermore, specific and non-specific inhibitors of several PDE protein families have been shown to be effective in treating such disorders. For example, the PDE4-specific inhibitor rolipram, mentioned above as an anti-depressant, inhibits lipopolysaccharide-induced expression of TNF- α ; and has been effective in treating multiple sclerosis in an animal model. Other PDE4-

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specific inhibitors are being investigated for use as anti-inflammatory therapeutics, and efficacy in attenuating the late asthmatic response to allergen challenge has been demonstrated [Harbinson, et al. (1997)]. Inhibitors specific for the PDE3 family have been approved for treatment of congestive heart failure. PDE5 inhibitors such as
5 Sildenafil are in use for treatment of penile erectile dysfunction [Terrett, N. et al. (1996)]. PDE5-inhibitors are under investigation as agents for cardiovascular therapy [Perry, M.J. and G.A. Higgs (1998)].

10 PDEs cyclic nucleotide levels have been suggested to influence proliferation of different cell types [Conti et al. (1995)]. For example, growth of the prostatic carcinoma cell lines DU145 and LNCaP was inhibited by cAMP derivatives and PDE inhibitors [Bang, Y.J. et al. (1994)]. Furthermore, PDEs have been implemented to additional cancers.

15 Non-specific inhibitors, such as theophylline and pentoxifylline, are currently used in the treatment of respiratory and vascular disorders, respectively.

In summary, cAMP and cGMP play a central role in intracellular second messenger signaling. Furthermore, the value as pharmaceutical targets has been proven for
20 several PDEs. Selective inhibitors have been developed as therapeutic agents for diseases such as cancer, heart failure, depression and sexual dysfunction. Thus, the identification of further disease implications of PDE species and their splice variants may lead to the development of specific inhibitors or modulators, or suggest new utilities for known compounds affecting PDEs. That in turn will provide additional
25 pharmacological approaches to treat diseases and conditions in which alterations in cyclic nucleotide pathways are involved. This diseases may include, but are not limited to, infections such as bacterial, fungal, protozoan, and viral infections, particularly those caused by HIV viruses, cancers, allergies including asthma, cardiovascular diseases including acute heart failure, hypotension, hypertension, angina pectoris, myocardial infarction, haematological diseases, genito-urinary
30 diseases including urinary incontinence and benign prostate hyperplasia,

osteoporosis, and peripheral and central nervous system disorders including pain, Alzheimer's disease and Parkinson's disease.

TaqMan-Technology / expression profiling

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TaqMan is a recently developed technique, in which the release of a fluorescent reporter dye from a hybridisation probe in real-time during a polymerase chain reaction (PCR) is proportional to the accumulation of the PCR product. Quantification is based on the early, linear part of the reaction, and by determining
10 the threshold cycle (CT), at which fluorescence above background is first detected.

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Gene expression technologies may be useful in several areas of drug discovery and development, such as target identification, lead optimization, and identification of mechanisms of action. The TaqMan technology can be used to compare differences
15 between expression profiles of normal tissue and diseased tissue. Expression profiling has been used in identifying genes, which are up- or downregulated in a variety of diseases. An interesting application of expression profiling is temporal monitoring of changes in gene expression during disease progression and drug treatment or in patients versus healthy individuals. The premise in this approach is
20 that changes in pattern of gene expression in response to physiological or environmental stimuli (e.g., drugs) may serve as indirect clues about disease-causing genes or drug targets. Moreover, the effects of drugs with established efficacy on global gene expression patterns may provide a guidepost, or a genetic signature, against which a new drug candidate can be compared.

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PDE11A

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The nucleotide sequence of PDE11A is accessible in public databases by the accession number NM_016953 and is given in SEQ ID NO:1. The amino acid sequence of PDE11A is depicted in SEQ ID NO:2.

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PDE11A is described as a phosphodiesterase [Fawcett L et al., (2000)]. The phosphodiesterase PDE11A is published in WO200040733 and WO200166716. The expression of PDE11A in prostate was previously described [Yuasa K et al. (2000)]. PDE11A shows the highest homology to other phosphodiesterases as shown in example 1.

Summary of the invention

The invention relates to novel disease associations of PDE11A polypeptides and polynucleotides. The invention also relates to novel methods of screening for therapeutic agents for the treatment of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal. The invention also relates to pharmaceutical compositions for the treatment of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising a PDE11A polypeptide, a PDE11A polynucleotide, or regulators of PDE11A or modulators of PDE11A activity. The invention further comprises methods of diagnosing disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal.

Brief Description of the Drawings

Fig. 1 shows the nucleotide sequence of a PDE11A polynucleotide (SEQ ID NO:1).

Fig. 2 shows the amino acid sequence of a PDE11A polypeptide (SEQ ID NO:2).

Fig. 3 shows the nucleotide sequence of a primer useful for the invention (SEQ ID NO:3).

Fig. 4 shows the nucleotide sequence of a primer useful for the invention (SEQ ID NO:4).

Fig. 5 shows a nucleotide sequence useful as a probe to detect proteins of the invention (SEQ ID NO:5).

Detailed description of the invention

Definition of terms

An "oligonucleotide" is a stretch of nucleotide residues which has a sufficient number of bases to be used as an oligomer, amplimer or probe in a polymerase chain reaction (PCR). Oligonucleotides are prepared from genomic or cDNA sequence and are used to amplify, reveal, or confirm the presence of a similar DNA or RNA in a particular cell or tissue. Oligonucleotides or oligomers comprise portions of a DNA sequence having at least about 10 nucleotides and as many as about 35 nucleotides, preferably about 25 nucleotides.

"Probes" may be derived from naturally occurring or recombinant single- or double-stranded nucleic acids or may be chemically synthesized. They are useful in detecting the presence of identical or similar sequences. Such probes may be labeled with reporter molecules using nick translation, Klenow fill-in reaction, PCR or other methods well known in the art. Nucleic acid probes may be used in southern, northern or in situ hybridizations to determine whether DNA or RNA encoding a certain protein is present in a cell type, tissue, or organ.

A "fragment of a polynucleotide" is a nucleic acid that comprises all or any part of a given nucleotide molecule, the fragment having fewer nucleotides than about 6 kb, preferably fewer than about 1 kb.

"Reporter molecules" are radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents which associate with a particular nucleotide or amino acid sequence, thereby establishing the presence of a certain sequence, or allowing for the quantification of a certain sequence.

“Chimeric” molecules may be constructed by introducing all or part of the nucleotide sequence of this invention into a vector containing additional nucleic acid sequence which might be expected to change any one or several of the following PDE11A characteristics: cellular location, distribution, ligand-binding affinities, interchain
5 affinities, degradation/turnover rate, signaling, etc.

“Active”, with respect to a PDE11A polypeptide, refers to those forms, fragments, or domains of a PDE11A polypeptide which retain the biological and/or antigenic
10 activity of a PDE11A polypeptide.

“Naturally occurring PDE11A polypeptide” refers to a polypeptide produced by cells which have not been genetically engineered and specifically contemplates various polypeptides arising from post-translational modifications of the polypeptide
15 including but not limited to acetylation, carboxylation, glycosylation, phosphorylation, lipidation and acylation.

“Derivative” refers to polypeptides which have been chemically modified by techniques such as ubiquitination, labeling (see above), pegylation (derivatization with polyethylene glycol), and chemical insertion or substitution of amino acids such
20 as ornithine which do not normally occur in human proteins.

“Conservative amino acid substitutions” result from replacing one amino acid with another having similar structural and/or chemical properties, such as the replacement
25 of a leucine with an isoleucine or valine, an aspartate with a glutamate, or a threonine with a serine.

“Insertions” or “deletions” are typically in the range of about 1 to 5 amino acids. The variation allowed may be experimentally determined by producing the peptide
30 synthetically while systematically making insertions, deletions, or substitutions of nucleotides in the sequence using recombinant DNA techniques.

5 A "signal sequence" or "leader sequence" can be used, when desired, to direct the polypeptide through a membrane of a cell. Such a sequence may be naturally present on the polypeptides of the present invention or provided from heterologous sources by recombinant DNA techniques.

10 An "oligopeptide" is a short stretch of amino acid residues and may be expressed from an oligonucleotide. Oligopeptides comprise a stretch of amino acid residues of at least 3, 5, 10 amino acids and at most 10, 15, 25 amino acids, typically of at least 9 to 13 amino acids, and of sufficient length to display biological and/or antigenic activity.

15 "Inhibitor" is any substance which retards or prevents a chemical or physiological reaction or response. Common inhibitors include but are not limited to antisense molecules, antibodies, and antagonists.

20 "Standard expression" is a quantitative or qualitative measurement for comparison. It is based on a statistically appropriate number of normal samples and is created to use as a basis of comparison when performing diagnostic assays, running clinical trials, or following patient treatment profiles.

25 "Animal" as used herein may be defined to include human, domestic (e.g., cats, dogs, etc.), agricultural (e.g., cows, horses, sheep, etc.) or test species (e.g., mouse, rat, rabbit, etc.).

A "PDE11A polynucleotide", within the meaning of the invention, shall be understood as being a nucleic acid molecule selected from a group consisting of

30 (i) nucleic acid molecules encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 2,

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- (ii) nucleic acid molecules comprising the sequence of SEQ ID NO: 1,
- (iii) nucleic acid molecules having the sequence of SEQ ID NO: 1,
- 5 (iv) nucleic acid molecules the complementary strand of which hybridizes under stringent conditions to a nucleic acid molecule of (i), (ii), or (iii); and
- (v) nucleic acid molecules the sequence of which differs from the sequence of a nucleic acid molecule of (iii) due to the degeneracy of the genetic code;

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wherein the polypeptide encoded by said nucleic acid molecule has PDE11A activity.

A "PDE11A polypeptide", within the meaning of the invention, shall be understood as being a polypeptide selected from a group consisting of

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- (i) polypeptides having the sequence of SEQ ID NO: 2,
- (ii) polypeptides comprising the sequence of SEQ ID NO: 2,
- 20 (iii) polypeptides encoded by PDE11A polynucleotides; and
- (iv) polypeptides which show at least 99%, 98%, 95%, 90%, or 80% homology with a polypeptide of (i), (ii), or (iii);

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wherein said polypeptide has PDE11A activity.

The nucleotide sequences encoding a PDE11A (or their complement) have numerous applications in techniques known to those skilled in the art of molecular biology. These techniques include use as hybridization probes, use in the construction of oligomers for PCR, use for chromosome and gene mapping, use in the recombinant production of PDE11A, and use in generation of antisense DNA or RNA, their

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chemical analogs and the like. Uses of nucleotides encoding a PDE11A disclosed herein are exemplary of known techniques and are not intended to limit their use in any technique known to a person of ordinary skill in the art. Furthermore, the nucleotide sequences disclosed herein may be used in molecular biology techniques that have not yet been developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, e.g., the triplet genetic code, specific base pair interactions, etc.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of PDE11A - encoding nucleotide sequences may be produced. Some of these will only bear minimal homology to the nucleotide sequence of the known and naturally occurring PDE11A. The invention has specifically contemplated each and every possible variation of nucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the nucleotide sequence of naturally occurring PDE11A, and all such variations are to be considered as being specifically disclosed.

Although the nucleotide sequences which encode a PDE11A, its derivatives or its variants are preferably capable of hybridizing to the nucleotide sequence of the naturally occurring PDE11A polynucleotide under stringent conditions, it may be advantageous to produce nucleotide sequences encoding PDE11A polypeptides or its derivatives possessing a substantially different codon usage. Codons can be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic expression host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding a PDE11A polypeptide and/or its derivatives without altering the encoded amino acid sequence include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

Nucleotide sequences encoding a PDE11A polypeptide may be joined to a variety of other nucleotide sequences by means of well established recombinant DNA techniques. Useful nucleotide sequences for joining to PDE11A polynucleotides include an assortment of cloning vectors such as plasmids, cosmids, lambda phage derivatives, phagemids, and the like. Vectors of interest include expression vectors, replication vectors, probe generation vectors, sequencing vectors, etc. In general, vectors of interest may contain an origin of replication functional in at least one organism, convenient restriction endonuclease sensitive sites, and selectable markers for one or more host cell systems.

Another aspect of the subject invention is to provide for PDE11A-specific hybridization probes capable of hybridizing with naturally occurring nucleotide sequences encoding PDE11A. Such probes may also be used for the detection of similar PDE encoding sequences and should preferably show at least 40% nucleotide identity to PDE11A polynucleotides. The hybridization probes of the subject invention may be derived from the nucleotide sequence presented as SEQ ID NO: 1 or from genomic sequences including promoter, enhancers or introns of the native gene. Hybridization probes may be labelled by a variety of reporter molecules using techniques well known in the art.

It will be recognized that many deletional or mutational analogs of PDE11A polynucleotides will be effective hybridization probes for PDE11A polynucleotides. Accordingly, the invention relates to nucleic acid sequences that hybridize with such PDE11A encoding nucleic acid sequences under stringent conditions.

"Stringent conditions" refers to conditions that allow for the hybridization of substantially related nucleic acid sequences. For instance, such conditions will generally allow hybridization of sequence with at least about 85% sequence identity, preferably with at least about 90% sequence identity, more preferably with at least about 95% sequence identity. Hybridization conditions and probes can be adjusted in well-characterized ways to achieve selective hybridization of human-derived

probes. Stringent conditions, within the meaning of the invention are 65°C in a buffer containing 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7 % (w/v) SDS.

5 Nucleic acid molecules that will hybridize to PDE11A polynucleotides under stringent conditions can be identified functionally. Without limitation, examples of the uses for hybridization probes include: histochemical uses such as identifying tissues that express PDE11A; measuring mRNA levels, for instance to identify a sample's tissue type or to identify cells that express abnormal levels of PDE11A; and detecting polymorphisms of PDE11A.

10 PCR provides additional uses for oligonucleotides based upon the nucleotide sequence which encodes PDE11A. Such probes used in PCR may be of recombinant origin, chemically synthesized, or a mixture of both. Oligomers may comprise discrete nucleotide sequences employed under optimized conditions for identification
15 of PDE11A in specific tissues or diagnostic use. The same two oligomers, a nested set of oligomers, or even a degenerate pool of oligomers may be employed under less stringent conditions for identification of closely related DNAs or RNAs.

20 Rules for designing polymerase chain reaction (PCR) primers are now established, as reviewed by PCR Protocols. Degenerate primers, i.e., preparations of primers that are heterogeneous at given sequence locations, can be designed to amplify nucleic acid sequences that are highly homologous to, but not identical with PDE11A. Strategies are now available that allow for only one of the primers to be required to specifically hybridize with a known sequence. For example, appropriate nucleic acid
25 primers can be ligated to the nucleic acid sought to be amplified to provide the hybridization partner for one of the primers. In this way, only one of the primers need be based on the sequence of the nucleic acid sought to be amplified.

30 PCR methods for amplifying nucleic acid will utilize at least two primers. One of these primers will be capable of hybridizing to a first strand of the nucleic acid to be amplified and of priming enzyme-driven nucleic acid synthesis in a first direction.

The other will be capable of hybridizing the reciprocal sequence of the first strand (if the sequence to be amplified is single stranded, this sequence will initially be hypothetical, but will be synthesized in the first amplification cycle) and of priming nucleic acid synthesis from that strand in the direction opposite the first direction and towards the site of hybridization for the first primer. Conditions for conducting such amplifications, particularly under preferred stringent hybridization conditions, are well known.

Other means of producing specific hybridization probes for PDE11A include the cloning of nucleic acid sequences encoding PDE11A or PDE11A derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerase as T7 or SP6 RNA polymerase and the appropriate reporter molecules.

It is possible to produce a DNA sequence, or portions thereof, entirely by synthetic chemistry. After synthesis, the nucleic acid sequence can be inserted into any of the many available DNA vectors and their respective host cells using techniques which are well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into the nucleotide sequence. Alternately, a portion of sequence in which a mutation is desired can be synthesized and recombined with longer portion of an existing genomic or recombinant sequence.

PDE11A polynucleotides may be used to produce a purified oligo-or polypeptide using well known methods of recombinant DNA technology. The oligopeptide may be expressed in a variety of host cells; either prokaryotic or eukaryotic. Host cells may be from the same species from which the nucleotide sequence was derived or from a different species. Advantages of producing an oligonucleotide by recombinant DNA technology include obtaining adequate amounts of the protein for purification and the availability of simplified purification procedures.

Quantitative determinations of nucleic acids

An important step in the molecular genetic analysis of human disease is often the enumeration of the copy number of a nucleic acid or the relative expression of a gene in particular tissues.

Several different approaches are currently available to make quantitative determinations of nucleic acids. Chromosome-based techniques, such as comparative genomic hybridization (CGH) and fluorescent in situ hybridization (FISH) facilitate efforts to cytogenetically localize genomic regions that are altered in tumor cells. Regions of genomic alteration can be narrowed further using loss of heterozygosity analysis (LOH), in which disease DNA is analyzed and compared with normal DNA for the loss of a heterozygous polymorphic marker. The first experiments used restriction fragment length polymorphisms (RFLPs) [Johnson, (1989)], or hypervariable minisatellite DNA [Barnes, 2000]. In recent years LOH has been performed primarily using PCR amplification of microsatellite markers and electrophoresis of the radio labelled [Jeffreys, (1985)] or fluorescently labelled PCR products [Weber, (1990)] and compared between paired normal and disease DNAs.

A number of other methods have also been developed to quantify nucleic acids [Gergen, (1992)]. More recently, PCR and RT-PCR methods have been developed which are capable of measuring the amount of a nucleic acid in a sample. One approach, for example, measures PCR product quantity in the log phase of the reaction before the formation of reaction products plateaus [Thomas, (1980)].

A gene sequence contained in all samples at relatively constant quantity is typically utilized for sample amplification efficiency normalization. This approach, however, suffers from several drawbacks. The method requires that each sample has equal input amounts of the nucleic acid and that the amplification efficiency between samples is identical until the time of analysis. Furthermore, it is difficult using the conventional methods of PCR quantitation such as gel electrophoresis or plate

capture hybridization to determine that all samples are in fact analyzed during the log phase of the reaction as required by the method.

Another method called quantitative competitive (QC)-PCR, as the name implies,
5 relies on the inclusion of an internal control competitor in each reaction [Piatak,
(1993), BioTechniques]. The efficiency of each reaction is normalized to the internal
competitor. A known amount of internal competitor is typically added to each
sample. The unknown target PCR product is compared with the known competitor
10 PCR product to obtain relative quantitation. A difficulty with this general approach
lies in developing an internal control that amplifies with the same efficiency than the
target molecule.

5' Fluorogenic Nuclease Assays

15 Fluorogenic nuclease assays are a real time quantitation method that uses a probe to
monitor formation of amplification product. The basis for this method of monitoring
the formation of amplification product is to measure continuously PCR product
accumulation using a dual-labelled fluorogenic oligonucleotide probe, an approach
frequently referred to in the literature simply as the "TaqMan method"
20 [Piatak,(1993), Science; Heid, (1996); Gibson, (1996); Holland. (1991)].

The probe used in such assays is typically a short (about 20-25 bases)
oligonucleotide that is labeled with two different fluorescent dyes. The 5' terminus
of the probe is attached to a reporter dye and the 3' terminus is attached to a
25 quenching dye, although the dyes could be attached at other locations on the probe as
well. The probe is designed to have at least substantial sequence complementarity
with the probe binding site. Upstream and downstream PCR primers which bind to
flanking regions of the locus are added to the reaction mixture. When the probe is
intact, energy transfer between the two fluorophors occurs and the quencher
30 quenches emission from the reporter. During the extension phase of PCR, the probe
is cleaved by the 5' nuclease activity of a nucleic acid polymerase such as Taq

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polymerase, thereby releasing the reporter from the oligonucleotide-quencher and resulting in an increase of reporter emission intensity which can be measured by an appropriate detector.

5 One detector which is specifically adapted for measuring fluorescence emissions such as those created during a fluorogenic assay is the ABI 7700 or 4700 HT manufactured by Applied Biosystems, Inc. in Foster City, Calif. The ABI 7700 uses fiber optics connected with each well in a 96-or 384 well PCR tube arrangement. The instrument includes a laser for exciting the labels and is capable of measuring
10 the fluorescence spectra intensity from each tube with continuous monitoring during PCR amplification. Each tube is re-examined every 8.5 seconds.

Computer software provided with the instrument is capable of recording the fluorescence intensity of reporter and quencher over the course of the amplification.
15 The recorded values will then be used to calculate the increase in normalized reporter emission intensity on a continuous basis. The increase in emission intensity is plotted versus time, i.e., the number of amplification cycles, to produce a continuous measure of amplification. To quantify the locus in each amplification reaction, the amplification plot is examined at a point during the log phase of product
20 accumulation. This is accomplished by assigning a fluorescence threshold intensity above background and determining the point at which each amplification plot crosses the threshold (defined as the threshold cycle number or Ct). Differences in threshold cycle number are used to quantify the relative amount of PCR target contained within each tube. Assuming that each reaction functions at 100% PCR efficiency, a
25 difference of one Ct represents a two-fold difference in the amount of starting template. The fluorescence value can be used in conjunction with a standard curve to determine the amount of amplification product present.

Non-Probe-Based Detection Methods

A variety of options are available for measuring the amplification products as they are formed. One method utilizes labels, such as dyes, which only bind to double stranded DNA. In this type of approach, amplification product (which is double stranded) binds dye molecules in solution to form a complex. With the appropriate dyes, it is possible to distinguish between dye molecules free in solution and dye molecules bound to amplification product. For example, certain dyes fluoresce only when bound to amplification product. Examples of dyes which can be used in methods of this general type include, but are not limited to, Syber Green.TM. and Pico Green from Molecular Probes, Inc. of Eugene, Oreg., ethidium bromide, propidium iodide, chromomycin, acridine orange, Hoechst 33258, Toto-1, Yoyo-1, DAPI (4',6-diamidino-2-phenylindole hydrochloride).

Another real time detection technique measures alteration in energy fluorescence energy transfer between fluorophors conjugated with PCR primers [Livak, (1995)].

Probe-Based Detection Methods

These detection methods involve some alteration to the structure or conformation of a probe hybridized to the locus between the amplification primer pair. In some instances, the alteration is caused by the template-dependent extension catalyzed by a nucleic acid polymerase during the amplification process. The alteration generates a detectable signal which is an indirect measure of the amount of amplification product formed.

For example, some methods involve the degradation or digestion of the probe during the extension reaction. These methods are a consequence of the 5'-3' nuclease activity associated with some nucleic acid polymerases. Polymerases having this activity cleave mononucleotides or small oligonucleotides from an oligonucleotide probe annealed to its complementary sequence located within the locus.

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The 3' end of the upstream primer provides the initial binding site for the nucleic acid polymerase. As the polymerase catalyzes extension of the upstream primer and encounters the bound probe, the nucleic acid polymerase displaces a portion of the 5' end of the probe and through its nuclease activity cleaves mononucleotides or oligonucleotides from the probe.

The upstream primer and the probe can be designed such that they anneal to the complementary strand in close proximity to one another. In fact, the 3' end of the upstream primer and the 5' end of the probe may abut one another. In this situation, extension of the upstream primer is not necessary in order for the nucleic acid polymerase to begin cleaving the probe. In the case in which intervening nucleotides separate the upstream primer and the probe, extension of the primer is necessary before the nucleic acid polymerase encounters the 5' end of the probe. Once contact occurs and polymerization continues, the 5'-3' exonuclease activity of the nucleic acid polymerase begins cleaving mononucleotides or oligonucleotides from the 5' end of the probe. Digestion of the probe continues until the remaining portion of the probe dissociates from the complementary strand.

In solution, the two end sections can hybridize with each other to form a hairpin loop. In this conformation, the reporter and quencher dye are in sufficiently close proximity that fluorescence from the reporter dye is effectively quenched by the quencher dye. Hybridized probe, in contrast, results in a linearized conformation in which the extent of quenching is decreased. Thus, by monitoring emission changes for the two dyes, it is possible to indirectly monitor the formation of amplification product.

Probes

The labeled probe is selected so that its sequence is substantially complementary to a segment of the test locus or a reference locus. As indicated above, the nucleic acid

site to which the probe binds should be located between the primer binding sites for the upstream and downstream amplification primers.

Primers

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The primers used in the amplification are selected so as to be capable of hybridizing to sequences at flanking regions of the locus being amplified. The primers are chosen to have at least substantial complementarity with the different strands of the nucleic acid being amplified. When a probe is utilized to detect the formation of amplification products, the primers are selected in such that they flank the probe, i.e. are located upstream and downstream of the probe.

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The primer must have sufficient length so that it is capable of priming the synthesis of extension products in the presence of an agent for polymerization. The length and composition of the primer depends on many parameters, including, for example, the temperature at which the annealing reaction is conducted, proximity of the probe binding site to that of the primer, relative concentrations of the primer and probe and the particular nucleic acid composition of the probe. Typically the primer includes 15-30 nucleotides. However, the length of the primer may be more or less depending on the complexity of the primer binding site and the factors listed above.

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Labels for Probes and Primers

The labels used for labeling the probes or primers of the current invention and which can provide the signal corresponding to the quantity of amplification product can take a variety of forms. As indicated above with regard to the 5' fluorogenic nuclease method, a fluorescent signal is one signal which can be measured. However, measurements may also be made, for example, by monitoring radioactivity, colorimetry, absorption, magnetic parameters, or enzymatic activity. Thus, labels which can be employed include, but are not limited to, fluorophors, chromophores,

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radioactive isotopes, electron dense reagents, enzymes, and ligands having specific binding partners (e.g., biotin-avidin).

Monitoring changes in fluorescence is a particularly useful way to monitor the accumulation of amplification products. A number of labels useful for attachment to probes or primers are commercially available including fluorescein and various fluorescein derivatives such as FAM, HEX, TET and JOE (all which are available from Applied Biosystems, Foster City, Calif.); lucifer yellow, and coumarin derivatives.

Labels may be attached to the probe or primer using a variety of techniques and can be attached at the 5' end, and/or the 3' end and/or at an internal nucleotide. The label can also be attached to spacer arms of various sizes which are attached to the probe or primer. These spacer arms are useful for obtaining a desired distance between multiple labels attached to the probe or primer.

In some instances, a single label may be utilized; whereas, in other instances, such as with the 5' fluorogenic nuclease assays for example, two or more labels are attached to the probe. In cases wherein the probe includes multiple labels, it is generally advisable to maintain spacing between the labels which is sufficient to permit separation of the labels during digestion of the probe through the 5'-3' nuclease activity of the nucleic acid polymerase.

Patients Exhibiting Symptoms of Disease

A number of diseases are associated with changes in the copy number of a certain gene. For patients having symptoms of a disease, the real-time PCR method can be used to determine if the patient has copy number alterations which are known to be linked with diseases that are associated with the symptoms the patient has.

PDE11A expression*PDE11A fusion proteins*

5 Fusion proteins are useful for generating antibodies against PDE11A polypeptides and for use in various assay systems. For example, fusion proteins can be used to identify proteins which interact with portions of PDE11A polypeptides. Protein affinity chromatography or library-based assays for protein-protein interactions, such as the yeast two-hybrid or phage display systems, can be used for this purpose. Such
10 methods are well known in the art and also can be used as drug screens.

A PDE11A fusion protein comprises two polypeptide segments fused together by means of a peptide bond. The first polypeptide segment can comprise at least 54, 75, 100, 125, 139, 150, 175, 200, 225, 250, 275, 300, 325 or 350 contiguous amino acids
15 of SEQ ID NO: 2 or of a biologically active variant, such as those described above. The first polypeptide segment also can comprise full-length PDE11A.

The second polypeptide segment can be a full-length protein or a protein fragment. Proteins commonly used in fusion protein construction include, but are not limited to
20 β galactosidase, β -glucuronidase, green fluorescent protein (GFP), autofluorescent proteins, including blue fluorescent protein (BFP), glutathione-S-transferase (GST), luciferase, horseradish peroxidase (HRP), and chloramphenicol acetyltransferase (CAT). Additionally, epitope tags are used in fusion protein constructions, including
25 histidine (His) tags, FLAG tags, influenza hemagglutinin (HA) tags, Myc tags, VSV-G tags, and thioredoxin (Trx) tags. Other fusion constructions can include maltose binding protein (MBP), S-tag, Lex a DNA binding domain (DBD) fusions, GAL4 DNA binding domain fusions, and herpes simplex virus (HSV) BP16 protein fusions. A fusion protein also can be engineered to contain a cleavage site located adjacent to the PDE11A.

Preparation of Polynucleotides

A naturally occurring PDE11A polynucleotide can be isolated free of other cellular components such as membrane components, proteins, and lipids. Polynucleotides
5 can be made by a cell and isolated using standard nucleic acid purification techniques, or synthesized using an amplification technique, such as the polymerase chain reaction (PCR), or by using an automatic synthesizer. Methods for isolating polynucleotides are routine and are known in the art. Any such technique for obtaining a polynucleotide can be used to obtain isolated PDE11A polynucleotides.
10 For example, restriction enzymes and probes can be used to isolate polynucleotide fragments which comprise PDE11A nucleotide sequences. Isolated polynucleotides are in preparations which are free or at least 70, 80, or 90% free of other molecules.

PDE11A cDNA molecules can be made with standard molecular biology techniques,
15 using PDE11A mRNA as a template. PDE11A cDNA molecules can thereafter be replicated using molecular biology techniques known in the art. An amplification technique, such as PCR, can be used to obtain additional copies of polynucleotides of the invention, using either human genomic DNA or cDNA as a template.

20 Alternatively, synthetic chemistry techniques can be used to synthesize PDE11A polynucleotides. The degeneracy of the genetic code allows alternate nucleotide sequences to be synthesized which will encode PDE11A having, for example, an amino acid sequence shown in SEQ ID NO: 2 or a biologically active variant thereof.

25 Extending Polynucleotides

Various PCR-based methods can be used to extend nucleic acid sequences encoding human PDE11A, for example to detect upstream sequences of PDE11A gene such as promoters and regulatory elements. For example, restriction-site PCR uses universal
30 primers to retrieve unknown sequence adjacent to a known locus. Genomic DNA is first amplified in the presence of a primer to a linker sequence and a primer specific

to the known region. The amplified sequences are then subjected to a second round of PCR with the same linker primer and another specific primer internal to the first one. Products of each round of PCR are transcribed with an appropriate RNA polymerase and sequenced using reverse transcriptase.

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Inverse PCR also can be used to amplify or extend sequences using divergent primers based on a known region. Primers can be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National Biosciences Inc., Plymouth, Minn.), to be 22-30 nucleotides in length, to have a GC content of 50% or more, and to anneal to the target sequence at temperatures about 68-72°C. The method uses several restriction enzymes to generate a suitable fragment in the known region of a gene. The fragment is then circularized by intramolecular ligation and used as a PCR template.

10

Another method which can be used is capture PCR, which involves PCR amplification of DNA fragments adjacent to a known sequence in human and yeast artificial chromosome DNA. In this method, multiple restriction enzyme digestions and ligations also can be used to place an engineered double-stranded sequence into an unknown fragment of the DNA molecule before performing PCR.

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When screening for full-length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. Randomly-primed libraries are preferable, in that they will contain more sequences which contain the 5' regions of genes. Use of a randomly primed library may be especially preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries can be useful for extension of sequence into 5' non-transcribed regulatory regions.

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Commercially available capillary electrophoresis systems can be used to analyze the size or confirm the nucleotide sequence of PCR or sequencing products. For example, capillary sequencing can employ flowable polymers for electrophoretic separation, four different fluorescent dyes (one for each nucleotide) which are laser

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activated, and detection of the emitted wavelengths by a charge coupled device camera. Output/light intensity can be converted to electrical signal using appropriate equipment and software (e.g., GENOTYPER and Sequence NAVIGATOR, Perkin Elmer), and the entire process from loading of samples to computer analysis and electronic data display can be computer controlled. Capillary electrophoresis is especially preferable for the sequencing of small pieces of DNA which might be present in limited amounts in a particular sample.

Obtaining Polypeptides

PDE11A can be obtained, for example, by purification from human cells, by expression of PDE11A polynucleotides, or by direct chemical synthesis.

Protein Purification

PDE11A can be purified from any human cell which expresses the enzyme, including those which have been transfected with expression constructs which express PDE11A. A purified PDE11A is separated from other compounds which normally associate with PDE11A in the cell, such as certain proteins, carbohydrates, or lipids, using methods well-known in the art. Such methods include, but are not limited to, size exclusion chromatography, ammonium sulfate fractionation, ion exchange chromatography, affinity chromatography, and preparative gel electrophoresis.

Expression of PDE11A Polynucleotides

To express PDE11A, PDE11A polynucleotides can be inserted into an expression vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art can be used to construct expression vectors containing sequences encoding PDE11A and appropriate transcriptional and translational control elements. These

methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination.

5 A variety of expression vector/host systems can be utilized to contain and express sequences encoding PDE11A. These include, but are not limited to, microorganisms, such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors, insect cell systems infected with virus expression vectors (*e.g.*, baculovirus), plant cell systems transformed with virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; 10 tobacco mosaic virus, TMV) or with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmids), or animal cell systems.

The control elements or regulatory sequences are those non-translated regions of the vector - enhancers, promoters, 5' and 3' untranslated regions - which interact with 15 host cellular proteins to carry out transcription and translation. Such elements can vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, can be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the 20 BLUESCRIPT phagemid (Stratagene, LaJolla, Calif.) or pSPORT1 plasmid (Life Technologies) and the like can be used. The baculovirus polyhedrin promoter can be used in insect cells. Promoters or enhancers derived from the genomes of plant cells (*e.g.*, heat shock, RUBISCO, and storage protein genes) or from plant viruses (*e.g.*, viral promoters or leader sequences) can be cloned into the vector. In mammalian 25 cell systems, promoters from mammalian genes or from mammalian viruses are preferable. If it is necessary to generate a cell line that contains multiple copies of a nucleotide sequence encoding PDE11A, vectors based on SV40 or EBV can be used with an appropriate selectable marker.

Bacterial and Yeast Expression Systems

In bacterial systems, a number of expression vectors can be selected. For example, when a large quantity of PDE11A is needed for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified can be used. Such vectors include, but are not limited to, multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene). In a BLUESCRIPT vector, a sequence encoding PDE11A can be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced. pIN vectors or pGEX vectors (Promega, Madison, Wis.) also can be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems can be designed to include heparin, thrombin, or factor Xa protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

Plant and Insect Expression Systems

If plant expression vectors are used, the expression of sequences encoding PDE11A can be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV can be used alone or in combination with the omega leader sequence from TMV. Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters can be used. These constructs can be introduced into plant cells by direct DNA transformation or by pathogen-mediated transfection.

An insect system also can be used to express PDE11A. For example, in one such system *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia*

larvae. Sequences encoding PDE11A can be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of PDE11A will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses can
5 then be used to infect *S. frugiperda* cells or *Trichoplusia* larvae in which PDE11A can be expressed.

Mammalian Expression Systems

10 A number of viral-based expression systems can be used to express PDE11A in mammalian host cells. For example, if an adenovirus is used as an expression vector, sequences encoding PDE11A can be ligated into an adenovirus transcription/-translation complex comprising the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome can be used to obtain
15 a viable virus which is capable of expressing PDE11A in infected host cells [Engelhard, 1994]. If desired, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, can be used to increase expression in mammalian host cells.

Human artificial chromosomes (HACs) also can be used to deliver larger fragments
20 of DNA than can be contained and expressed in a plasmid. HACs of 6M to 10M are constructed and delivered to cells via conventional delivery methods (*e.g.*, liposomes, polycationic amino polymers, or vesicles). Specific initiation signals also can be used to achieve more efficient translation of sequences encoding PDE11A. Such signals include the ATG initiation codon and adjacent sequences. In cases where
25 sequences encoding PDE11A, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals (including the ATG initiation codon) should be provided. The initiation codon
30 should be in the correct reading frame to ensure translation of the entire insert.

Exogenous translational elements and initiation codons can be of various origins, both natural and synthetic.

Host Cells

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A host cell strain can be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed PDE11A in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the polypeptide also can be
10 used to facilitate correct insertion, folding and/or function. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (*e.g.*, CHO, HeLa, MDCK, HEK293, and WI38), are available from the American Type Culture Collection (ATCC; 10801 University Boulevard, Manassas,
15 VA 20110-2209) and can be chosen to ensure the correct modification and processing of the foreign protein.

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Stable expression is preferred for long-term, high-yield production of recombinant proteins. For example, cell lines which stably express PDE11A can be transformed using expression vectors which can contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells can be allowed to grow for 1-2 days in an enriched medium before they are switched to a selective medium. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced PDE11A sequences. Resistant clones of stably transformed cells can be proliferated using tissue culture techniques appropriate to the cell type. Any number of selection systems can be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase [Logan, (1984)] and adenine phosphoribosyltransferase [Wigler, (1977)] genes which can be employed in *tk* or *aprt* cells, respectively. Also, antimetabolite, antibiotic, or herbicide resistance

can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate [Lowy, (1980)], *npt* confers resistance to the aminoglycosides, neomycin and G-418 [Wigler, (1980)], and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively [Colbere-Garapin, 5 1981]. Additional selectable genes have been described. For example, *trpB* allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine. Visible markers such as anthocyanins, β -glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, can be used to identify transformants and to quantify the amount of transient or stable protein 10 expression attributable to a specific vector system

Detecting Polypeptide Expression

Although the presence of marker gene expression suggests that a PDE11A polynucleotide is also present, its presence and expression may need to be confirmed. 15 For example, if a sequence encoding PDE11A is inserted within a marker gene sequence, transformed cells containing sequences which encode PDE11A can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding PDE11A under the control of a single 20 promoter. Expression of the marker gene in response to induction or selection usually indicates expression of PDE11A polynucleotide.

Alternatively, host cells which contain a PDE11A polynucleotide and which express PDE11A can be identified by a variety of procedures known to those of skill in the 25 art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques which include membrane, solution, or chip-based technologies for the detection and/or quantification of nucleic acid or protein. For example, the presence of a polynucleotide sequence encoding PDE11A can be detected by DNA-DNA or DNA- 30 RNA hybridization or amplification using probes or fragments or fragments of polynucleotides encoding PDE11A. Nucleic acid amplification-based assays involve

the use of oligonucleotides selected from sequences encoding PDE11A to detect transformants which contain a PDE11A polynucleotide.

5 A variety of protocols for detecting and measuring the expression of PDE11A, using either polyclonal or monoclonal antibodies specific for the polypeptide, are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay using monoclonal antibodies reactive to two non-interfering epitopes on PDE11A can be used, or a competitive binding assay can
10 be employed.

A wide variety of labels and conjugation techniques are known by those skilled in the art and can be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to
15 polynucleotides encoding PDE11A include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, sequences encoding PDE11A can be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and can be used to synthesize RNA probes *in vitro* by addition of labeled nucleotides and an appropriate
20 RNA polymerase such as T7, T3, or SP6. These procedures can be conducted using a variety of commercially available kits (Amersham Pharmacia Biotech, Promega, and US Biochemical). Suitable reporter molecules or labels which can be used for ease of detection include radionuclides, enzymes, and fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles,
25 and the like.

Expression and Purification of Polypeptides

Host cells transformed with PDE11A polynucleotides can be cultured under
30 conditions suitable for the expression and recovery of the protein from cell culture. The polypeptide produced by a transformed cell can be secreted or contained

intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing PDE11A polynucleotides can be designed to contain signal sequences which direct secretion of soluble PDE11A through a prokaryotic or eukaryotic cell membrane or which direct the membrane insertion of membrane-bound PDE11A.

As discussed above, other constructions can be used to join a sequence encoding PDE11A to a nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). Inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, CA) between the purification domain and PDE11A also can be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing PDE11A and 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification by IMAC (immobilized metal ion affinity chromatography) Maddox, (1983)], while the enterokinase cleavage site provides a means for purifying PDE11A from the fusion protein [Porath, (1992)].

Chemical Synthesis

Sequences encoding PDE11A can be synthesized, in whole or in part, using chemical methods well known in the art. Alternatively, PDE11A itself can be produced using chemical methods to synthesize its amino acid sequence, such as by direct peptide synthesis using solid-phase techniques. Protein synthesis can either be performed using manual techniques or by automation. Automated synthesis can be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer).

Optionally, fragments of PDE11A can be separately synthesized and combined using chemical methods to produce a full-length molecule.

5 The newly synthesized peptide can be substantially purified by preparative high performance liquid chromatography. The composition of a synthetic PDE11A can be confirmed by amino acid analysis or sequencing. Additionally, any portion of the amino acid sequence of PDE11A can be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins to produce a variant polypeptide or a fusion protein.

10 *Production of Altered Polypeptides*

As will be understood by those of skill in the art, it may be advantageous to produce PDE11A polynucleotides possessing non-naturally occurring codons. For example, 15 codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce an RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

20 The nucleotide sequences referred to herein can be engineered using methods generally known in the art to alter PDE11A polynucleotides for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the polypeptide or mRNA product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides 25 can be used to engineer the nucleotide sequences. For example, site-directed mutagenesis can be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, introduce mutations, and so forth.

Antibodies

Any type of antibody known in the art can be generated to bind specifically to an epitope of PDE11A.

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“Antibody” as used herein includes intact immunoglobulin molecules, as well as fragments thereof, such as Fab, F(ab')₂, and Fv, which are capable of binding an epitope of PDE11A. Typically, at least 6, 8, 10, or 12 contiguous amino acids are required to form an epitope. However, epitopes which involve non-contiguous amino acids may require more, *e.g.*, at least 15, 25, or 50 amino acid. An antibody which specifically binds to an epitope of PDE11A can be used therapeutically, as well as in immunochemical assays, such as Western blots, ELISAs, radioimmunoassays, immunohistochemical assays, immunoprecipitations, or other immunochemical assays known in the art. Various immunoassays can be used to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays are well known in the art. Such immunoassays typically involve the measurement of complex formation between an immunogen and an antibody which specifically binds to the PDE11A immunogen.

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Typically, an antibody which specifically binds to PDE11A provides a detection signal at least 5-, 10-, or 20-fold higher than a detection signal provided with other proteins when used in an immunochemical assay. Preferably, antibodies which specifically bind to PDE11A do not detect other proteins in immunochemical assays and can immunoprecipitate PDE11A from solution.

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PDE11A can be used to immunize a mammal, such as a mouse, rat, rabbit, guinea pig, monkey, or human, to produce polyclonal antibodies. If desired, PDE11A can be conjugated to a carrier protein, such as bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin. Depending on the host species, various adjuvants can be used to increase the immunological response. Such adjuvants include, but are not limited to, Freund's adjuvant, mineral gels (*e.g.*, aluminum hydroxide), and surface

active substances (e.g., lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and dinitrophenol). Among adjuvants used in humans, BCG (*bacilli Calmette-Guerin*) and *Corynebacterium parvum* are especially useful.

5

Monoclonal antibodies which specifically bind to PDE11A can be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These techniques include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-
10 hybridoma technique [Roberge, (1995)].

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In addition, techniques developed for the production of "chimeric antibodies", the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. Monoclonal and other antibodies also can be "humanized" to prevent a patient from mounting an immune response against the antibody when it is used therapeutically. Such antibodies may be sufficiently similar in sequence to human antibodies to be used directly in therapy or may require alteration of a few key residues. Sequence differences between rodent antibodies and human sequences can be minimized by
20 replacing residues which differ from those in the human sequences by site directed mutagenesis of individual residues or by grating of entire complementarity determining regions. Antibodies which specifically bind to PDE11A can contain antigen binding sites which are either partially or fully humanized, as disclosed in U.S. 5,565,332.

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Alternatively, techniques described for the production of single chain antibodies can be adapted using methods known in the art to produce single chain antibodies which specifically bind to PDE11A. Antibodies with related specificity, but of distinct idiotypic composition, can be generated by chain shuffling from random
30 combinatorial immunoglobulin libraries. Single-chain antibodies also can be constructed using a DNA amplification method, such as PCR, using hybridoma

cDNA as a template. Single-chain antibodies can be mono- or bispecific, and can be bivalent or tetravalent. Construction of tetravalent, bispecific single-chain antibodies is taught. A nucleotide sequence encoding a single-chain antibody can be constructed using manual or automated nucleotide synthesis, cloned into an expression construct using standard recombinant DNA methods, and introduced into a cell to express the coding sequence, as described below. Alternatively, single-chain antibodies can be produced directly using, for example, filamentous phage technology.

Antibodies which specifically bind to PDE11A also can be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents. Other types of antibodies can be constructed and used therapeutically in methods of the invention. For example, chimeric antibodies can be constructed as disclosed in WO 93/03151. Binding proteins which are derived from immunoglobulins and which are multivalent and multispecific, such as the "diabodies" described in WO 94/13804, also can be prepared.

Antibodies according to the invention can be purified by methods well known in the art. For example, antibodies can be affinity purified by passage over a column to which PDE11A is bound. The bound antibodies can then be eluted from the column using a buffer with a high salt concentration.

Antisense Oligonucleotides

Antisense oligonucleotides are nucleotide sequences which are complementary to a specific DNA or RNA sequence. Once introduced into a cell, the complementary nucleotides combine with natural sequences produced by the cell to form complexes and block either transcription or translation. Preferably, an antisense oligonucleotide is at least 11 nucleotides in length, but can be at least 12, 15, 20, 25, 30, 35, 40, 45, or 50 or more nucleotides long. Longer sequences also can be used. Antisense

- 38 -

oligonucleotide molecules can be provided in a DNA construct and introduced into a cell as described above to decrease the level of PDE11A gene products in the cell.

Antisense oligonucleotides can be deoxyribonucleotides, ribonucleotides, or a combination of both. Oligonucleotides can be synthesized manually or by an automated synthesizer, by covalently linking the 5' end of one nucleotide with the 3' end of another nucleotide with non-phosphodiester internucleotide linkages such as alkylphosphonates, phosphorothioates, phosphorodithioates, alkylphosphonothioates, alkylphosphonates, phosphoramidates, phosphate esters, carbamates, acetamides, carboxymethyl esters, carbonates, and phosphate triesters.

Modifications of PDE11A gene expression can be obtained by designing antisense oligonucleotides which will form duplexes to the control, 5', or regulatory regions of the PDE11A gene. Oligonucleotides derived from the transcription initiation site, *e.g.*, between positions -10 and +10 from the start site, are preferred. Similarly, inhibition can be achieved using "triple helix" base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or chaperons. Therapeutic advances using triplex DNA have been described in the literature [Nicholls, (1993)]. An antisense oligonucleotide also can be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Precise complementarity is not required for successful complex formation between an antisense oligonucleotide and the complementary sequence of a PDE11A polynucleotide. Antisense oligonucleotides which comprise, for example, 2, 3, 4, or 5 or more stretches of contiguous nucleotides which are precisely complementary to a PDE11A polynucleotide, each separated by a stretch of contiguous nucleotides which are not complementary to adjacent PDE11A nucleotides, can provide sufficient targeting specificity for PDE11A mRNA. Preferably, each stretch of complementary contiguous nucleotides is at least 4, 5, 6, 7, or 8 or more nucleotides in length. Non-complementary intervening sequences are preferably 1, 2, 3, or 4 nucleotides in

length. One skilled in the art can easily use the calculated melting point of an antisense-sense pair to determine the degree of mismatching which will be tolerated between a particular antisense oligonucleotide and a particular PDE11A polynucleotide sequence. Antisense oligonucleotides can be modified without
5 affecting their ability to hybridize to a PDE11A polynucleotide. These modifications can be internal or at one or both ends of the antisense molecule. For example, internucleoside phosphate linkages can be modified by adding cholesteryl or diamine moieties with varying numbers of carbon residues between the amino groups and terminal ribose. Modified bases and/or sugars, such as arabinose instead of ribose, or
10 a 3', 5'-substituted oligonucleotide in which the 3' hydroxyl group or the 5' phosphate group are substituted, also can be employed in a modified antisense oligonucleotide. These modified oligonucleotides can be prepared by methods well known in the art.

Ribozymes

15 Ribozymes are RNA molecules with catalytic activity [Uhlmann, (1987)]. Ribozymes can be used to inhibit gene function by cleaving an RNA sequence, as is known in the art. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by
20 endonucleolytic cleavage. Examples include engineered hammerhead motif ribozyme molecules that can specifically and efficiently catalyze endonucleolytic cleavage of specific nucleotide sequences. The coding sequence of a PDE11A polynucleotide can be used to generate ribozymes which will specifically bind to mRNA transcribed from a PDE11A polynucleotide. Methods of designing and
25 constructing ribozymes which can cleave other RNA molecules in trans in a highly sequence specific manner have been developed and described in the art. For example, the cleavage activity of ribozymes can be targeted to specific RNAs by engineering a discrete "hybridization" region into the ribozyme. The hybridization region contains a sequence complementary to the target RNA and thus specifically
30 hybridizes with the target RNA.

Specific ribozyme cleavage sites within a PDE11A RNA target can be identified by scanning the target molecule for ribozyme cleavage sites which include the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target RNA
5 containing the cleavage site can be evaluated for secondary structural features which may render the target inoperable. Suitability of candidate PDE11A RNA targets also can be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays. The nucleotide sequences shown in SEQ ID NO: 1 and its complement provide sources of suitable
10 hybridization region sequences. Longer complementary sequences can be used to increase the affinity of the hybridization sequence for the target. The hybridizing and cleavage regions of the ribozyme can be integrally related such that upon hybridizing to the target RNA through the complementary regions, the catalytic region of the ribozyme can cleave the target.

15 Ribozymes can be introduced into cells as part of a DNA construct. Mechanical methods, such as microinjection, liposome-mediated transfection, electroporation, or calcium phosphate precipitation, can be used to introduce a ribozyme-containing DNA construct into cells in which it is desired to decrease PDE11A expression.
20 Alternatively, if it is desired that the cells stably retain the DNA construct, the construct can be supplied on a plasmid and maintained as a separate element or integrated into the genome of the cells, as is known in the art. A ribozyme-encoding DNA construct can include transcriptional regulatory elements, such as a promoter element, an enhancer or UAS element, and a transcriptional terminator signal, for
25 controlling transcription of ribozymes in the cells (U.S. 5,641,673). Ribozymes also can be engineered to provide an additional level of regulation, so that destruction of mRNA occurs only when both a ribozyme and a target gene are induced in the cells.

Screening / Screening Assays*Regulators*

5 Regulators as used herein, refer to compounds that affect the activity of PDE11A in vivo and/or in vitro. Regulators can be agonists and antagonists of PDE11A polypeptide and can be compounds that exert their effect on the PDE11A activity via the enzymatic activity, expression, post-translational modifications or by other means. Agonists of PDE11A are molecules which, when bound to PDE11A, increase or prolong the activity of PDE11A. Agonists of PDE11A include proteins, nucleic acids, carbohydrates, small molecules, or any other molecule which activate PDE11A. Antagonists of PDE11A are molecules which, when bound to PDE11A, decrease the amount or the duration of the activity of PDE11A. Antagonists include proteins, nucleic acids, carbohydrates, antibodies, small molecules, or any other molecule which decrease the activity of PDE11A.

The term "modulate", as it appears herein, refers to a change in the activity of PDE11A polypeptide. For example, modulation may cause an increase or a decrease in enzymatic activity, binding characteristics, or any other biological, functional, or immunological properties of PDE11A.

As used herein, the terms "specific binding" or "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, or an antagonist. The interaction is dependent upon the presence of a particular structure of the protein recognized by the binding molecule (i.e., the antigenic determinant or epitope). For example, if an antibody is specific for epitope "A" the presence of a polypeptide containing the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The invention provides methods (also referred to herein as "screening assays") for identifying compounds which can be used for the treatment of hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-uological disorders and inflammation diseases. The methods entail the identification of candidate or test compounds or agents (e.g., peptides, peptidomimetics, small molecules or other molecules) which bind to PDE11A and/or have a stimulatory or inhibitory effect on the biological activity of PDE11A or its expression and then determining which of these compounds have an effect on symptoms or diseases regarding the hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-uological disorders and inflammation diseases in an *in vivo* assay.

Candidate or test compounds or agents which bind to PDE11A and/or have a stimulatory or inhibitory effect on the activity or the expression of PDE11A are identified either in assays that employ cells which express PDE11A (cell-based assays) or in assays with isolated PDE11A (cell-free assays). The various assays can employ a variety of variants of PDE11A (e.g., full-length PDE11A, a biologically active fragment of PDE11A, or a fusion protein which includes all or a portion of PDE11A). Moreover, PDE11A can be derived from any suitable mammalian species (e.g., human PDE11A, rat PDE11A or murine PDE11A). The assay can be a binding assay entailing direct or indirect measurement of the binding of a test compound or a known PDE11A ligand to PDE11A. The assay can also be an activity assay entailing direct or indirect measurement of the activity of PDE11A. The assay can also be an expression assay entailing direct or indirect measurement of the expression of PDE11A mRNA or PDE11A protein. The various screening assays are combined with an *in vivo* assay entailing measuring the effect of the test compound on the symptoms of hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-uological disorders and inflammation diseases.

The present invention includes biochemical, cell free assays that allow the identification of inhibitors and agonists of PDEs suitable as lead structures for pharmacological drug development. Such assays involve contacting a form of PDE11A (e.g., full-length PDE11A, a biologically active fragment of PDE11A, or a fusion protein comprising all or a portion of PDE11A) with a test compound and determining the ability of the test compound to act as an antagonist (preferably) or an agonist of the enzymatic activity of PDE11A. In one embodiment, the assay includes monitoring the PDE activity of PDE11A by measuring the conversion of either cAMP or cGMP to its nucleoside monophosphate after contacting PDE11A with a test compound.

For example, cAMP and cGMP levels can be measured by the use of the tritium containing compounds $^3\text{HcAMP}$ and $^3\text{HcGMP}$ as described in [Hansen, R.S., and Beavo, J.A., PNAS USA1982;79: 2788-92]. To screen a compound pool comprised of a large number of compounds, the microtiter plate-based scintillation proximity assay (SPA) as described in [Bardelle, C. et al. (1999) Anal. Biochem. 275: 148-155] can be applied.

Alternatively, the phosphodiesterase activity of the recombinant protein can be assayed using a commercially available SPA kit (Amersham Pharmacia). The PDE enzyme hydrolyzes cyclic nucleotides, e.g. cAMP and cGMP to their linear counterparts. The SPA assay utilizes the tritiated cyclic nucleotides [^3H]cAMP or [^3H]cGMP, and is based upon the selective interaction of the tritiated non cyclic product with the SPA beads whereas the cyclic substrates are not effectively binding. Radiolabelled product bound to the scintillation beads generates light that can be analyzed in a scintillation counter.

In one embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of PDE11A. Such assays can employ full-length PDE11A, a biologically active fragment of PDE11A, or a fusion protein which includes all or a portion of PDE11A. As described in greater detail

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below, the test compound can be obtained by any suitable means, e.g., from conventional compound libraries.

5 Determining the ability of the test compound to modulate the activity of PDE11A can be accomplished, for example, by determining the ability of PDE11A to bind to or interact with a target molecule. The target molecule can be a molecule with which PDE11A binds or interacts with in nature. The target molecule can be a component of a signal transduction pathway which facilitates transduction of an extracellular signal. The target PDE11A molecule can be, for example, a second intracellular
10 protein which has catalytic activity or a protein which facilitates the association of downstream signaling molecules with PDE11A.

Determining the ability of PDE11A to bind to or interact with a target molecule can be accomplished by one of the methods described above for determining direct
15 binding. In one embodiment, determining the ability of a polypeptide of the invention to bind to or interact with a target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (e.g., intracellular Ca^{2+} , diacylglycerol, IP_3 , etc.), detecting
20 catalytic/enzymatic activity of the target on an appropriate substrate, detecting the induction of a reporter gene (e.g., a regulatory element that is responsive to a polypeptide of the invention operably linked to a nucleic acid encoding a detectable marker, e.g., luciferase), or detecting a cellular response.

25 In various embodiments of the above assay methods of the present invention, it may be desirable to immobilize PDE11A (or a PDE11A target molecule) to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to PDE11A, or interaction of PDE11A with a target molecule in the presence and
30 absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test

tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase (GST) fusion proteins or glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical; St. Louis, Mo.) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or PDE11A, and the mixture incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components and complex formation is measured either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of binding or activity of PDE11A can be determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either PDE11A or its target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated polypeptide of the invention or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals; Rockford, Ill.), and immobilized in the wells of streptavidin-coated plates (Pierce Chemical). Alternatively, antibodies reactive with PDE11A or target molecules but which do not interfere with binding of the polypeptide of the invention to its target molecule can be derivatized to the wells of the plate, and unbound target or polypeptide of the invention trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with PDE11A or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with PDE11A or target molecule.

Another technique for drug screening which may be used provides for high throughput screening of compounds having suitable binding affinity to the protein of interest as described in published PCT application WO84/03564. In this method, large numbers of different small test compounds are synthesized on a solid substrate, such as plastic pins or some other surface. The test compounds are reacted with PDE11A, or fragments thereof, and washed. Bound PDE11A is then detected by methods well known in the art. Purified PDE11A can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding PDE11A specifically compete with a test compound for binding PDE11A. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with PDE11A.

The screening assay can also involve monitoring the expression of PDE11A. For example, regulators of expression of PDE11A can be identified in a method in which a cell is contacted with a candidate compound and the expression of PDE11A protein or mRNA in the cell is determined. The level of expression of PDE11A protein or mRNA the presence of the candidate compound is compared to the level of expression of PDE11A protein or mRNA in the absence of the candidate compound. The candidate compound can then be identified as a regulator of expression of PDE11A based on this comparison. For example, when expression of PDE11A protein or mRNA protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of PDE11A protein or mRNA expression. Alternatively, when expression of PDE11A protein or mRNA is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of PDE11A protein or mRNA expression. The level of

PDE11A protein or mRNA expression in the cells can be determined by methods described below.

Binding Assays

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For binding assays, the test compound is preferably a small molecule which binds to and occupies the active site of PDE11A polypeptide, thereby making the ligand binding site inaccessible to substrate such that normal biological activity is prevented. Examples of such small molecules include, but are not limited to, small peptides or peptide-like molecules. Potential ligands which bind to a polypeptide of the invention include, but are not limited to, the natural ligands of known PDE11A PDEs and analogues or derivatives thereof.

15

In binding assays, either the test compound or the PDE11A polypeptide can comprise a detectable label, such as a fluorescent, radioisotopic, chemiluminescent, or enzymatic label, such as horseradish peroxidase, alkaline phosphatase, or luciferase. Detection of a test compound which is bound to PDE11A polypeptide can then be accomplished, for example, by direct counting of radioemmission, by scintillation counting, or by determining conversion of an appropriate substrate to a detectable product. Alternatively, binding of a test compound to a PDE11A polypeptide can be determined without labeling either of the interactants. For example, a microphysiometer can be used to detect binding of a test compound with a PDE11A polypeptide. A microphysiometer (*e.g.*, Cytosensor™) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between a test compound and PDE11A [Haseloff, (1988)].

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Determining the ability of a test compound to bind to PDE11A also can be accomplished using a technology such as real-time Bimolecular Interaction Analysis (BIA) [McConnell, (1992); Sjolander, (1991)]. BIA is a technology for studying

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biospecific interactions in real time, without labeling any of the interactants (*e.g.*, BIAcore™). Changes in the optical phenomenon surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

5 In yet another aspect of the invention, a PDE11A-like polypeptide can be used as a "bait protein" in a two-hybrid assay or three-hybrid assay [Szabo, (1995); U.S. 5,283,317], to identify other proteins which bind to or interact with PDE11A and modulate its activity.

10 The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. For example, in one construct, polynucleotide encoding PDE11A can be fused to a polynucleotide encoding the DNA binding domain of a known transcription factor (*e.g.*, GAL-4). In the other construct a DNA
15 sequence that encodes an unidentified protein ("prey" or "sample") can be fused to a polynucleotide that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact *in vivo* to form an protein-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a
20 reporter gene (*e.g.*, LacZ), which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected, and cell colonies containing the functional transcription factor can be isolated and used to obtain the DNA sequence encoding the protein which interacts with PDE11A.

25 It may be desirable to immobilize either the PDE11A (or polynucleotide) or the test compound to facilitate separation of the bound form from unbound forms of one or both of the interactants, as well as to accommodate automation of the assay. Thus, either the PDE11A-like polypeptide (or polynucleotide) or the test compound can be
30 bound to a solid support. Suitable solid supports include, but are not limited to, glass or plastic slides, tissue culture plates, microtiter wells, tubes, silicon chips, or

particles such as beads (including, but not limited to, latex, polystyrene, or glass beads). Any method known in the art can be used to attach PDE11A-like polypeptide (or polynucleotide) or test compound to a solid support, including use of covalent and non-covalent linkages, passive absorption, or pairs of binding moieties attached respectively to the polypeptide (or polynucleotide) or test compound and the solid support. Test compounds are preferably bound to the solid support in an array, so that the location of individual test compounds can be tracked. Binding of a test compound to PDE11A (or a polynucleotide encoding for PDE11A) can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtiter plates, test tubes, and microcentrifuge tubes.

In one embodiment, PDE11A is a fusion protein comprising a domain that allows binding of PDE11A to a solid support. For example, glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, Mo.) or glutathione derivatized microtiter plates, which are then combined with the test compound or the test compound and the non-adsorbed PDE11A; the mixture is then incubated under conditions conducive to complex formation (*e.g.*, at physiological conditions for salt and pH). Following incubation, the beads or microtiter plate wells are washed to remove any unbound components. Binding of the interactants can be determined either directly or indirectly, as described above. Alternatively, the complexes can be dissociated from the solid support before binding is determined.

Other techniques for immobilizing proteins or polynucleotides on a solid support also can be used in the screening assays of the invention. For example, either PDE11A (or a polynucleotide encoding PDE11A) or a test compound can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated PDE11A (or a polynucleotide encoding biotinylated PDE11A) or test compounds can be prepared from biotin-NHS (N-hydroxysuccinimide) using techniques well known in the art (*e.g.*, biotinylation kit, Pierce Chemicals, Rockford, Ill.) and immobilized in the wells of streptavidin-coated plates (Pierce Chemical). Alternatively, antibodies

which specifically bind to PDE11A, polynucleotide, or a test compound, but which do not interfere with a desired binding site, such as the active site of PDE11A, can be derivatized to the wells of the plate. Unbound target or protein can be trapped in the wells by antibody conjugation.

5

Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies which specifically bind to PDE11A polypeptide or test compound, enzyme-linked assays which rely on detecting an activity of PDE11A polypeptide, and SDS gel electrophoresis under non-reducing conditions.

10

Screening for test compounds which bind to a PDE11A polypeptide or polynucleotide also can be carried out in an intact cell. Any cell which comprises a PDE11A polypeptide or polynucleotide can be used in a cell-based assay system. A PDE11A polynucleotide can be naturally occurring in the cell or can be introduced using techniques such as those described above. Binding of the test compound to PDE11A or a polynucleotide encoding PDE11A is determined as described above.

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Functional Assays

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Test compounds can be tested for the ability to increase or decrease PDE11A activity of a PDE11A polypeptide. The PDE11A activity can be measured, for example, using methods described in the specific examples, below. PDE11A activity can be measured after contacting either a purified PDE11A or an intact cell with a test compound. A test compound which decreases PDE11A activity by at least about 10, preferably about 50, more preferably about 75, 90, or 100% is identified as a potential agent for decreasing PDE11A activity. A test compound which increases PDE11A activity by at least about 10, preferably about 50, more preferably about 75, 90, or 100% is identified as a potential agent for increasing PDE11A activity.

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Gene Expression

In another embodiment, test compounds which increase or decrease PDE11A gene expression are identified. As used herein, the term "correlates with expression of a polynucleotide" indicates that the detection of the presence of nucleic acids, the same or related to a nucleic acid sequence encoding PDE11A, by northern analysis or realtime PCR is indicative of the presence of nucleic acids encoding PDE11A in a sample, and thereby correlates with expression of the transcript from the polynucleotide encoding PDE11A. The term "microarray", as used herein, refers to an array of distinct polynucleotides or oligonucleotides arrayed on a substrate, such as paper, nylon or any other type of membrane, filter, chip, glass slide, or any other suitable solid support. A PDE11A polynucleotide is contacted with a test compound, and the expression of an RNA or polypeptide product of PDE11A polynucleotide is determined. The level of expression of appropriate mRNA or polypeptide in the presence of the test compound is compared to the level of expression of mRNA or polypeptide in the absence of the test compound. The test compound can then be identified as a regulator of expression based on this comparison. For example, when expression of mRNA or polypeptide is greater in the presence of the test compound than in its absence, the test compound is identified as a stimulator or enhancer of the mRNA or polypeptide expression. Alternatively, when expression of the mRNA or polypeptide is less in the presence of the test compound than in its absence, the test compound is identified as an inhibitor of the mRNA or polypeptide expression.

The level of PDE11A mRNA or polypeptide expression in the cells can be determined by methods well known in the art for detecting mRNA or polypeptide. Either qualitative or quantitative methods can be used. The presence of polypeptide products of PDE11A polynucleotide can be determined, for example, using a variety of techniques known in the art, including immunochemical methods such as radioimmunoassay, Western blotting, and immunohistochemistry. Alternatively, polypeptide synthesis can be determined *in vivo*, in a cell culture, or in an *in vitro* translation system by detecting incorporation of labelled amino acids into PDE11A.

Such screening can be carried out either in a cell-free assay system or in an intact cell. Any cell which expresses PDE11A polynucleotide can be used in a cell-based assay system. The PDE11A polynucleotide can be naturally occurring in the cell or
5 can be introduced using techniques such as those described above. Either a primary culture or an established cell line can be used.

Test Compounds

10 Suitable test compounds for use in the screening assays of the invention can be obtained from any suitable source, e.g., conventional compound libraries. The test compounds can also be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries;
15 spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of
20 compounds [Lam, (1997)]. Examples of methods for the synthesis of molecular libraries can be found in the art. Libraries of compounds may be presented in solution or on beads, bacteria, spores, plasmids or phage.

Modeling of Regulators

25 Computer modeling and searching technologies permit identification of compounds, or the improvement of already identified compounds, that can modulate PDE11A expression or activity. Having identified such a compound or composition, the active sites or regions are identified. Such sites might typically be the enzymatic active site, regulator binding sites, or ligand binding sites. The active site can be identified using
30 methods known in the art including, for example, from the amino acid sequences of peptides, from the nucleotide sequences of nucleic acids, or from study of complexes

of the relevant compound or composition with its natural ligand. In the latter case, chemical or X-ray crystallographic methods can be used to find the active site by finding where on the factor the complexed ligand is found.

5 Next, the three dimensional geometric structure of the active site is determined. This can be done by known methods, including X-ray crystallography, which can determine a complete molecular structure. On the other hand, solid or liquid phase NMR can be used to determine certain intramolecular distances. Any other
10 experimental method of structure determination can be used to obtain partial or complete geometric structures. The geometric structures may be measured with a complexed ligand, natural or artificial, which may increase the accuracy of the active site structure determined.

If an incomplete or insufficiently accurate structure is determined, the methods of
15 computer based numerical modeling can be used to complete the structure or improve its accuracy. Any recognized modeling method may be used, including parameterized models specific to particular biopolymers such as proteins or nucleic acids, molecular dynamics models based on computing molecular motions, statistical mechanics models based on thermal ensembles, or combined models. For most types
20 of models, standard molecular force fields, representing the forces between constituent atoms and groups, are necessary, and can be selected from force fields known in physical chemistry. The incomplete or less accurate experimental structures can serve as constraints on the complete and more accurate structures computed by these modeling methods.

25 Finally, having determined the structure of the active site, either experimentally, by modeling, or by a combination, candidate modulating compounds can be identified by searching databases containing compounds along with information on their molecular structure. Such a search seeks compounds having structures that match the
30 determined active site structure and that interact with the groups defining the active

site. Such a search can be manual, but is preferably computer assisted. These compounds found from this search are potential PDE11A modulating compounds.

Alternatively, these methods can be used to identify improved modulating compounds from an already known modulating compound or ligand. The composition of the known compound can be modified and the structural effects of modification can be determined using the experimental and computer modeling methods described above applied to the new composition. The altered structure is then compared to the active site structure of the compound to determine if an improved fit or interaction results. In this manner systematic variations in composition, such as by varying side groups, can be quickly evaluated to obtain modified modulating compounds or ligands of improved specificity or activity.

Therapeutic Indications and Methods

It was found by the present applicant that PDE11A is expressed in various human tissues.

Central Nervous System (CNS) Disorders

CNS disorders include disorders of the central nervous system as well as disorders of the peripheral nervous system.

CNS disorders include, but are not limited to brain injuries, cerebrovascular diseases and their consequences, Parkinson's disease, corticobasal degeneration, motor neuron disease, dementia, including ALS, „mild cognitive impairment“, age-associated loss of learning and memory, attention-deficit disorder, multiple sclerosis, traumatic brain injury, stroke, post-stroke, post-traumatic brain injury, and small-vessel cerebrovascular disease. Dementias, such as Alzheimer's disease, vascular dementia, dementia with Lewy bodies, frontotemporal dementia and Parkinsonism linked to chromosome 17, frontotemporal dementias, including Pick's disease, progressive

nuclear palsy, corticobasal degeneration, Huntington's disease, thalamic degeneration, Creutzfeld-Jakob dementia, HIV dementia, schizophrenia with dementia, and Korsakoff's psychosis, within the meaning of the definition are also considered to be CNS disorders.

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Similarly, cognitive-related disorders, such as mild cognitive impairment, age-associated memory impairment, age-related cognitive decline, vascular cognitive impairment, attention deficit disorders, attention deficit hyperactivity disorders, and memory disturbances in children with learning disabilities are also considered to be

10 CNS disorders.

15

Pain, within the meaning of this definition, is also considered to be a CNS disorder. Pain can be associated with CNS disorders, such as multiple sclerosis, spinal cord injury, sciatica, failed back surgery syndrome, traumatic brain injury, epilepsy, Parkinson's disease, post-stroke, and vascular lesions in the brain and spinal cord (e.g., infarct, hemorrhage, vascular malformation). Non-central neuropathic pain includes that associated with post mastectomy pain, phantom feeling, reflex sympathetic dystrophy (RSD), trigeminal neuralgia, radioculopathy, post-surgical pain, HIV/AIDS related pain, cancer pain, metabolic neuropathies (e.g., diabetic neuropathy, vasculitic neuropathy secondary to connective tissue disease), paraneoplastic polyneuropathy associated, for example, with carcinoma of lung, or leukemia, or lymphoma, or carcinoma of prostate, colon or stomach, trigeminal neuralgia, cranial neuralgias, and post-herpetic neuralgia. Pain associated with peripheral nerve damage, central pain (i.e. due to cerebral ischemia) and various

20 chronic pain i.e., lumbago, back pain (low back pain), inflammatory and/or rheumatic pain. Headache pain (for example, migraine with aura, migraine without aura, and other migraine disorders), episodic and chronic tension-type headache, tension-type like headache, cluster headache, and chronic paroxysmal hemicrania are also CNS disorders.

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Visceral pain such as pancreatitis, intestinal cystitis, dysmenorrhea, irritable Bowel syndrome, Crohn's disease, biliary colic, ureteral colic, myocardial infarction and pain syndromes of the pelvic cavity, e.g., vulvodynia, orchialgia, urethral syndrome and protatodynia are also CNS disorders.

5

Also considered to be a disorder of the nervous system are acute pain, for example postoperative pain, and pain after trauma.

10

The human PDE11A is highly expressed in the following brain tissues: fetal brain, Alzheimer cerebral cortex, thalamus, cerebral cortex. The expression in brain tissues and in particular the differential expression between diseased tissue Alzheimer cerebral cortex and healthy tissue cerebral cortex demonstrates that the human PDE11A or mRNA can be utilized to diagnose nervous system diseases. Additionally the activity of the human PDE11A can be modulated to treat nervous system diseases.

15

Cardiovascular Disorders

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Heart failure is defined as a pathophysiological state in which an abnormality of cardiac function is responsible for the failure of the heart to pump blood at a rate commensurate with the requirement of the metabolizing tissue. It includes all forms of pumping failures such as high-output and low-output, acute and chronic, right-sided or left-sided, systolic or diastolic, independent of the underlying cause.

25

Myocardial infarction (MI) is generally caused by an abrupt decrease in coronary blood flow that follows a thrombotic occlusion of a coronary artery previously narrowed by arteriosclerosis. MI prophylaxis (primary and secondary prevention) is included as well as the acute treatment of MI and the prevention of complications.

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Ischemic diseases are conditions in which the coronary flow is restricted resulting in a perfusion which is inadequate to meet the myocardial requirement for oxygen. This

group of diseases includes stable angina, unstable angina and asymptomatic ischemia.

5 Arrhythmias include all forms of atrial and ventricular tachyarrhythmias, atrial tachycardia, atrial flutter, atrial fibrillation, atrio-ventricular reentrant tachycardia, preexcitation syndrome, ventricular tachycardia, ventricular flutter, ventricular fibrillation, as well as bradycardic forms of arrhythmias.

10 Hypertensive vascular diseases include primary as well as all kinds of secondary arterial hypertension, renal, endocrine, neurogenic, others. The genes may be used as drug targets for the treatment of hypertension as well as for the prevention of all complications arising from cardiovascular diseases.

15 Peripheral vascular diseases are defined as vascular diseases in which arterial and/or venous flow is reduced resulting in an imbalance between blood supply and tissue oxygen demand. It includes chronic peripheral arterial occlusive disease (PAOD), acute arterial thrombosis and embolism, inflammatory vascular disorders, Raynaud's phenomenon and venous disorders.

20 Atherosclerosis is a cardiovascular disease in which the vessel wall is remodeled, compromising the lumen of the vessel. The atherosclerotic remodeling process involves accumulation of cells, both smooth muscle cells and monocyte/macrophage inflammatory cells, in the intima of the vessel wall. These cells take up lipid, likely from the circulation, to form a mature atherosclerotic lesion. Although the formation
25 of these lesions is a chronic process, occurring over decades of an adult human life, the majority of the morbidity associated with atherosclerosis occurs when a lesion ruptures, releasing thrombogenic debris that rapidly occludes the artery. When such an acute event occurs in the coronary artery, myocardial infarction can ensue, and in the worst case, can result in death.

The formation of the atherosclerotic lesion can be considered to occur in five overlapping stages such as migration, lipid accumulation, recruitment of inflammatory cells, proliferation of vascular smooth muscle cells, and extracellular matrix deposition. Each of these processes can be shown to occur in man and in
5 animal models of atherosclerosis, but the relative contribution of each to the pathology and clinical significance of the lesion is unclear.

Thus, a need exists for therapeutic methods and agents to treat cardiovascular pathologies, such as atherosclerosis and other conditions related to coronary artery
10 disease.

Cardiovascular diseases include but are not limited to disorders of the heart and the vascular system like congestive heart failure, myocardial infarction, ischemic diseases of the heart, all kinds of atrial and ventricular arrhythmias, hypertensive
15 vascular diseases, peripheral vascular diseases, and atherosclerosis.

The human PDE11A is highly expressed in the following cardiovascular related tissues: heart ventricle (left), pericardium, interventricular septum. Expression in the above mentioned tissues demonstrates that the human PDE11A or mRNA can be
20 utilized to diagnose of cardiovascular diseases. Additionally the activity of the human PDE11A can be modulated to treat cardiovascular diseases.

Genitourological Disorders

Genitourological disorders comprise benign and malign disorders of the organs constituting the genitourological system of female and male, renal diseases like acute or chronic renal failure, immunologically mediated renal diseases like renal transplant rejection, lupus nephritis, immune complex renal diseases, glomerulopathies, nephritis, toxic nephropathy, obstructive uropathies like benign prostatic hyperplasia (BPH), neurogenic bladder syndrome, urinary incontinence like urge-, stress-, or overflow incontinence, pelvic pain, and erectile dysfunction.

The human PDE11A is highly expressed in the following tissues of the genito-urinary system: testis, prostate. The expression in the above mentioned tissues demonstrates that the human PDE11A or mRNA can be utilized to diagnose of genito-urinary disorders. Additionally the activity of the human PDE11A can be modulated to treat genito-urinary disorders.

Liver Diseases

Liver diseases comprise primary or secondary, acute or chronic diseases or injury of the liver which may be acquired or inherited, benign or malignant, and which may affect the liver or the body as a whole. They comprise but are not limited to disorders of the bilirubin metabolism, jaundice, syndroms of Gilbert's, Crigler-Najjar, Dubin-Johnson and Rotor; intrahepatic cholestasis, hepatomegaly, portal hypertension, ascites, Budd-Chiari syndrome, portal-systemic encephalopathy, fatty liver, steatosis, Reye's syndrome, liver diseases due to alcohol, alcoholic hepatitis or cirrhosis, fibrosis and cirrhosis, fibrosis and cirrhosis of the liver due to inborn errors of metabolism or exogenous substances, storage diseases, syndromes of Gaucher's, Zellweger's, Wilson's - disease, acute or chronic hepatitis, viral hepatitis and its variants, inflammatory conditions of the liver due to viruses, bacteria, fungi, protozoa, helminths; drug induced disorders of the liver, chronic liver diseases like primary sclerosing cholangitis, α_1 -antitrypsin-deficiency, primary biliary

cirrhosis, postoperative liver disorders like postoperative intrahepatic cholestasis, hepatic granulomas, vascular liver disorders associated with systemic disease, benign or malignant neoplasms of the liver, disturbance of liver metabolism in the new-born or prematurely born.

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The human PDE11A is highly expressed in the following liver tissues: liver, liver liver cirrhosis. The expression in liver tissues and in particular the differential expression between diseased tissue liver liver cirrhosis and healthy tissue liver demonstrates that the human PDE11A or mRNA can be utilized to diagnose of liver diseases. Additionally the activity of the human PDE11A can be modulated to treat those diseases.

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Cancer Disorders

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Cancer disorders within the scope of this definition comprise any disease of an organ or tissue in mammals characterized by poorly controlled or uncontrolled multiplication of normal or abnormal cells in that tissue and its effect on the body as a whole. Cancer diseases within the scope of the definition comprise benign neoplasms, dysplasias, hyperplasias as well as neoplasms showing metastatic growth or any other transformations like e.g. leukoplakias which often precede a breakout of cancer. Cells and tissues are cancerous when they grow more rapidly than normal cells, displacing or spreading into the surrounding healthy tissue or any other tissues of the body described as metastatic growth, assume abnormal shapes and sizes, show changes in their nucleocytoplasmatic ratio, nuclear polychromasia, and finally may cease. Cancerous cells and tissues may affect the body as a whole when causing paraneoplastic syndromes or if cancer occurs within a vital organ or tissue, normal function will be impaired or halted, with possible fatal results. The ultimate involvement of a vital organ by cancer, either primary or metastatic, may lead to the death of the mammal affected. Cancer tends to spread, and the extent of its spread is usually related to an individual's chances of surviving the disease. Cancers are generally said to be in one of three stages of growth: early, or localized, when a

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tumor is still confined to the tissue of origin, or primary site; direct extension, where cancer cells from the tumour have invaded adjacent tissue or have spread only to regional lymph nodes; or metastasis, in which cancer cells have migrated to distant parts of the body from the primary site, via the blood or lymph systems, and have established secondary sites of infection. Cancer is said to be malignant because of its tendency to cause death if not treated. Benign tumors usually do not cause death, although they may if they interfere with a normal body function by virtue of their location, size, or paraneoplastic side effects. Hence benign tumors fall under the definition of cancer within the scope of this definition as well. In general, cancer cells divide at a higher rate than do normal cells, but the distinction between the growth of cancerous and normal tissues is not so much the rapidity of cell division in the former as it is the partial or complete loss of growth restraint in cancer cells and their failure to differentiate into a useful, limited tissue of the type that characterizes the functional equilibrium of growth of normal tissue. Cancer tissues may express certain molecular receptors and probably are influenced by the host's susceptibility and immunity and it is known that certain cancers of the breast and prostate, for example, are considered dependent on specific hormones for their existence. The term "cancer" under the scope of the definition is not limited to simple benign neoplasia but comprises any other benign and malign neoplasia like 1) Carcinoma, 2) Sarcoma, 3) Carcinosarcoma, 4) Cancers of the blood-forming tissues, 5) tumors of nerve tissues including the brain, 6) cancer of skin cells. Cancer according to 1) occurs in epithelial tissues, which cover the outer body (the skin) and line mucous membranes and the inner cavitory structures of organs e.g. such as the breast, lung, the respiratory and gastrointestinal tracts, the endocrine glands, and the genitourinary system. Ductal or glandular elements may persist in epithelial tumors, as in adenocarcinomas like e.g. thyroid adenocarcinoma, gastric adenocarcinoma, uterine adenocarcinoma. Cancers of the pavement-cell epithelium of the skin and of certain mucous membranes, such as e.g. cancers of the tongue, lip, larynx, urinary bladder, uterine cervix, or penis, may be termed epidermoid or squamous-cell carcinomas of the respective tissues and are in the scope of the definition of cancer as well. Cancer according to 2) develops in connective tissues, including fibrous tissues, adipose (fat)

tissues, muscle, blood vessels, bone, and cartilage like e.g. osteogenic sarcoma; liposarcoma, fibrosarcoma, synovial sarcoma. Cancer according to 3) is cancer that develops in both epithelial and connective tissue. Cancer disease within the scope of this definition may be primary or secondary, whereby primary indicates that the cancer originated in the tissue where it is found rather than was established as a secondary site through metastasis from another lesion. Cancers and tumor diseases within the scope of this definition may be benign or malign and may affect all anatomical structures of the body of a mammal. By example but not limited to they comprise cancers and tumor diseases of I) the bone marrow and bone marrow derived cells (leukemias), II) the endocrine and exocrine glands like e.g. thyroid, parathyroid, pituitary, adrenal glands, salivary glands, pancreas III) the breast, like e.g. benign or malignant tumors in the mammary glands of either a male or a female, the mammary ducts, adenocarcinoma, medullary carcinoma, comedo carcinoma, Paget's disease of the nipple, inflammatory carcinoma of the young woman, IV) the lung, V) the stomach, VI) the liver and spleen, VII) the small intestine, VIII) the colon, IX) the bone and its supportive and connective tissues like malignant or benign bone tumour, e.g. malignant osteogenic sarcoma, benign osteoma, cartilage tumors; like malignant chondrosarcoma or benign chondroma; bone marrow tumors like malignant myeloma or benign eosinophilic granuloma, as well as metastatic tumors from bone tissues at other locations of the body; X) the mouth, throat, larynx, and the esophagus, XI) the urinary bladder and the internal and external organs and structures of the urogenital system of male and female like ovaries, uterus, cervix of the uterus, testes, and prostate gland, XII) the prostate, XIII) the pancreas, like ductal carcinoma of the pancreas; XIV) the lymphatic tissue like lymphomas and other tumors of lymphoid origin, XV) the skin, XVI) cancers and tumor diseases of all anatomical structures belonging to the respiration and respiratory systems including thoracal muscles and linings, XVII) primary or secondary cancer of the lymph nodes XVIII) the tongue and of the bony structures of the hard palate or sinuses, XVIV) the mouth, cheeks, neck and salivary glands, XX) the blood vessels including the heart and their linings, XXI) the smooth or skeletal muscles and their ligaments and linings, XXII) the

peripheral, the autonomous, the central nervous system including the cerebellum, XXIII) the adipose tissue.

5 The human PDE11A is highly expressed in the following cancer tissues: thyroid tumor, lung tumor, breast tumor, colon tumor. The expression in the above mentioned tissues and in particular the differential expression between diseased tissue thyroid tumor and healthy tissue thyroid, between diseased tissue lung tumor and healthy tissue lung, between diseased tissue breast tumor and healthy tissue breast, between diseased tissue colon tumor and healthy tissue colon demonstrates
10 that the human PDE11A or mRNA can be utilized to diagnose of cancer. Additionally the activity of the human PDE11A can be modulated to treat cancer.

Applications

15 The present invention provides for both prophylactic and therapeutic methods for disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases.

20 The regulatory method of the invention involves contacting a cell with an agent that modulates one or more of the activities of PDE11A. An agent that modulates activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring cognate ligand of the polypeptide, a peptide, a peptidomimetic, or any small molecule. In one embodiment, the agent stimulates one or more of the biological activities of PDE11A. Examples of such stimulatory agents include the
25 active PDE11A and nucleic acid molecules encoding a portion of PDE11A. In another embodiment, the agent inhibits one or more of the biological activities of PDE11A. Examples of such inhibitory agents include antisense nucleic acid molecules and antibodies. These regulatory methods can be performed in vitro (e.g., by culturing the cell with the agent) or, alternatively, in vivo (e.g., by administering
30 the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by unwanted

expression or activity of PDE11A or a protein in the PDE11A signaling pathway. In one embodiment, the method involves administering an agent like any agent identified or being identifiable by a screening assay as described herein, or combination of such agents that modulate say upregulate or downregulate the expression or activity of PDE11A or of any protein in the PDE11A signaling pathway. In another embodiment, the method involves administering a regulator of PDE11A as therapy to compensate for reduced or undesirably low expression or activity of PDE11A or a protein in the PDE11A signaling pathway.

Stimulation of activity or expression of PDE11A is desirable in situations in which enzymatic activity or expression is abnormally low and in which increased activity is likely to have a beneficial effect. Conversely, inhibition of enzymatic activity or expression of PDE11A is desirable in situations in which activity or expression of PDE11A is abnormally high and in which decreasing its activity is likely to have a beneficial effect.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application are hereby incorporated by reference.

Pharmaceutical Compositions

This invention further pertains to novel agents identified by the above-described screening assays and uses thereof for treatments as described herein.

The nucleic acid molecules, polypeptides, and antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal

agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof
5 in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

The invention includes pharmaceutical compositions comprising a regulator of PDE11A expression or activity (and/or a regulator of the activity or expression of a
10 protein in the PDE11A signaling pathway) as well as methods for preparing such compositions by combining one or more such regulators and a pharmaceutically acceptable carrier. Also within the invention are pharmaceutical compositions comprising a regulator identified using the screening assays of the invention packaged with instructions for use. For regulators that are antagonists of PDE11A
15 activity or which reduce PDE11A expression, the instructions would specify use of the pharmaceutical composition for treatment of hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-urological disorders and inflammation diseases. For regulators that are agonists of PDE11A activity or increase PDE11A expression, the instructions would
20 specify use of the pharmaceutical composition for treatment of hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-urological disorders and inflammation diseases.

An inhibitor of PDE11A may be produced using methods which are generally known
25 in the art. In particular, purified PDE11A may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind PDE11A. Antibodies to PDE11A may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, single chain antibodies, Fab fragments, and fragments
30 produced by a Fab expression library. Neutralizing antibodies like those which inhibit dimer formation are especially preferred for therapeutic use.

In another embodiment of the invention, the polynucleotides encoding PDE11A, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotide encoding PDE11A may be used in
5 situations in which it would be desirable to block the transcription of the mRNA. In particular, cells may be transformed with sequences complementary to polynucleotides encoding PDE11A. Thus, complementary molecules or fragments may be used to modulate PDE11A activity, or to achieve regulation of gene function. Such technology is now well known in the art, and sense or antisense
10 oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding PDE11A.

Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide
15 sequences to the targeted organ, tissue, or cell population. Methods which are well known to those skilled in the art can be used to construct vectors which will express nucleic acid sequence complementary to the polynucleotides of the gene encoding PDE11A. These techniques are described, for example, in [Scott and Smith (1990)].

20 Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

An additional embodiment of the invention relates to the administration of a
25 pharmaceutical composition containing PDE11A in conjunction with a pharmaceutically acceptable carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of PDE11A, antibodies to PDE11A, and mimetics, agonists, antagonists, or inhibitors of PDE11A. The compositions may be administered alone or in combination with at least one other
30 agent, such as a stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier including, but not limited to, saline, buffered

saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs or hormones.

5 A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline
10 solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be
15 adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

20 Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor EMTM (BASF, Parsippany, N.J.) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be
25 fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, a pharmaceutically acceptable polyol like glycerol, propylene glycol, liquid
30 polyethylene glycol, and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance

of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic
5 agents, for example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin. Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a polypeptide or antibody) in
10 the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the
15 preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be
20 enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed.

25 Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient
30 such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or sterotes; a glidant such as

colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

5 For administration by inhalation, the compounds are delivered in the form of an aerosol spray from a pressurized container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

10 Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally
15 known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

20 In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid.
25 Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as
30 pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration. For pharmaceutical compositions which include an antagonist of PDE11A activity, a compound which reduces expression of PDE11A, or a compound which reduces expression or activity of a protein in the PDE11A signaling pathway or any combination thereof, the instructions for administration will specify use of the composition for hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-urological disorders and inflammation diseases. For pharmaceutical compositions which include an agonist of PDE11A activity, a compound which increases expression of PDE11A, or a compound which increases expression or activity of a protein in the PDE11A signaling pathway or any combination thereof, the instructions for administration will specify use of the composition for hematological and cardiovascular diseases, disorders of the peripheral and central nervous system, COPD, asthma, genito-urological disorders and inflammation diseases.

Diagnostics

5 In another embodiment, antibodies which specifically bind PDE11A may be used for the diagnosis of disorders characterized by the expression of PDE11A, or in assays to monitor patients being treated with PDE11A or agonists, antagonists, and inhibitors of PDE11A. Antibodies useful for diagnostic purposes may be prepared in the same manner as those described above for therapeutics. Diagnostic assays for PDE11A include methods which utilize the antibody and a label to detect PDE11A in human body fluids or in extracts of cells or tissues. The antibodies may be used with or
10 without modification, and may be labeled by covalent or non-covalent joining with a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

15 A variety of protocols for measuring PDE11A, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of PDE11A expression. Normal or standard values for PDE11A expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, preferably human, with antibody to PDE11A under conditions suitable for complex formation. The amount of standard complex formation may be quantified by
20 various methods, preferably by photometric means. Quantities of PDE11A expressed in subject samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

25 In another embodiment of the invention, the polynucleotides encoding PDE11A may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantitate gene expression in biopsied tissues in which expression of PDE11A may be correlated with disease. The
30 diagnostic assay may be used to distinguish between absence, presence, and excess

expression of PDE11A, and to monitor regulation of PDE11A levels during therapeutic intervention.

5 Polynucleotide sequences encoding PDE11A may be used for the diagnosis of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases associated with expression of PDE11A. The polynucleotide sequences encoding PDE11A may be used in Southern, Northern, or dot-blot analysis, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and ELISA assays; and in microarrays utilizing
10 fluids or tissues from patient biopsies to detect altered PDE11A expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding PDE11A may be useful in assays that detect the presence of associated disorders, particularly those mentioned
15 above. The nucleotide sequences encoding PDE11A may be labelled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantitated and compared with a standard value. If the amount of signal in the patient sample is significantly altered
20 from that of a comparable control sample, the nucleotide sequences have hybridized with nucleotide sequences in the sample, and the presence of altered levels of nucleotide sequences encoding PDE11A in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or in
25 monitoring the treatment of an individual patient.

In order to provide a basis for the diagnosis of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases associated with expression of PDE11A, a normal or standard profile for
30 expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or

a fragment thereof, encoding PDE11A, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained from normal samples may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Determination of a Therapeutically Effective Dose

The determination of a therapeutically effective dose is well within the capability of those skilled in the art. A therapeutically effective dose refers to that amount of active ingredient which increases or decreases PDE11A activity relative to PDE11A activity which occurs in the absence of the therapeutically effective dose. For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays or in animal models, usually mice, rabbits, dogs, or pigs. The animal model also can be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

Therapeutic efficacy and toxicity, e.g., ED_{50} (the dose therapeutically effective in 50% of the population) and LD_{50} (the dose lethal to 50% of the population), can be determined by standard pharmaceutical procedures in cell cultures or experimental animals. The dose ratio of toxic to therapeutic effects is the therapeutic index, and it can be expressed as the ratio, LD_{50}/ED_{50} . Pharmaceutical compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies is used in formulating a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that include the ED_{50} with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, sensitivity of the patient, and the route of administration. The exact dosage will be determined by the

- practitioner, in light of factors related to the subject that requires treatment. Dosage and administration are adjusted to provide sufficient levels of the active ingredient or to maintain the desired effect. Factors which can be taken into account include the severity of the disease state, general health of the subject, age, weight, and gender of the subject, diet, time and frequency of administration, drug combination(s), reaction sensitivities, and tolerance/response to therapy. Long-acting pharmaceutical compositions can be administered every 3 to 4 days, every week, or once every two weeks depending on the half-life and clearance rate of the particular formulation.
- Normal dosage amounts can vary from 0.1 micrograms to 100,000 micrograms, up to a total dose of about 1 g, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc. If the reagent is a single-chain antibody, polynucleotides encoding the antibody can be constructed and introduced into a cell either *ex vivo* or *in vivo* using well-established techniques including, but not limited to, transferrin-polycation-mediated DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated cellular fusion, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, "gene gun", and DEAE- or calcium phosphate-mediated transfection.
- If the expression product is mRNA, the reagent is preferably an antisense oligonucleotide or a ribozyme. Polynucleotides which express antisense oligonucleotides or ribozymes can be introduced into cells by a variety of methods, as described above. Preferably, a reagent reduces expression of PDE11A gene or the activity of PDE11A by at least about 10, preferably about 50, more preferably about 75, 90, or 100% relative to the absence of the reagent. The effectiveness of the mechanism chosen to decrease the level of expression of PDE11A gene or the activity of PDE11A can be assessed using methods well known in the art, such as hybridization

of nucleotide probes to PDE11A-specific mRNA, quantitative RT-PCR, immunologic detection of PDE11A, or measurement of PDE11A activity.

5 In any of the embodiments described above, any of the pharmaceutical compositions of the invention can be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy can be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents can act synergistically to effect the treatment or prevention of the various disorders described
10 above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects. Any of the therapeutic methods described above can be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

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Nucleic acid molecules of the invention are those nucleic acid molecules which are contained in a group of nucleic acid molecules consisting of (i) nucleic acid molecules encoding a polypeptide comprising the amino acid sequence of SEQ ID
20 NO: 2, (ii) nucleic acid molecules comprising the sequence of SEQ ID NO: 1, (iii) nucleic acid molecules having the sequence of SEQ ID NO: 1, (iv) nucleic acid molecules the complementary strand of which hybridizes under stringent conditions to a nucleic acid molecule of (i), (ii), or (iii); and (v) nucleic acid molecules the sequence of which differs from the sequence of a nucleic acid molecule of (iii) due to
25 the degeneracy of the genetic code, wherein the polypeptide encoded by said nucleic acid molecule has PDE11A activity.

Polypeptides of the invention are those polypeptides which are contained in a group of polypeptides consisting of (i) polypeptides having the sequence of SEQ ID NO: 2,
30 (ii) polypeptides comprising the sequence of SEQ ID NO: 2, (iii) polypeptides encoded by nucleic acid molecules of the invention and (iv) polypeptides which

show at least 99%, 98%, 95%, 90%, or 80% homology with a polypeptide of (i), (ii), or (iii), wherein said purified polypeptide has PDE11A activity.

5 An object of the invention is a method of screening for therapeutic agents useful in the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising the steps of (i) contacting a test compound with a PDE11A polypeptide, (ii) detect binding of said test compound to said PDE11A polypeptide. E.g., compounds that bind to the
10 PDE11A polypeptide are identified potential therapeutic agents for such a disease.

Another object of the invention is a method of screening for therapeutic agents useful in the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver
15 disease and genito-urinary diseases in a mammal comprising the steps of (i) determining the activity of a PDE11A polypeptide at a certain concentration of a test compound or in the absence of said test compound, (ii) determining the activity of said polypeptide at a different concentration of said test compound. E.g., compounds that lead to a difference in the activity of the PDE11A polypeptide in (i) and (ii) are
20 identified potential therapeutic agents for such a disease.

Another object of the invention is a method of screening for therapeutic agents useful in the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver
25 disease and genito-urinary diseases in a mammal comprising the steps of (i) determining the activity of a PDE11A polypeptide at a certain concentration of a test compound, (ii) determining the activity of a PDE11A polypeptide at the presence of a compound known to be a regulator of a PDE11A polypeptide. E.g., compounds that show similar effects on the activity of the PDE11A polypeptide in (i) as
30 compared to compounds used in (ii) are identified potential therapeutic agents for such a disease.

Other objects of the invention are methods of the above, wherein the step of contacting is in or at the surface of a cell.

5 Other objects of the invention are methods of the above, wherein the cell is in vitro.

Other objects of the invention are methods of the above, wherein the step of contacting is in a cell-free system.

10 Other objects of the invention are methods of the above, wherein the polypeptide is coupled to a detectable label.

Other objects of the invention are methods of the above, wherein the compound is coupled to a detectable label.

15 Other objects of the invention are methods of the above, wherein the test compound displaces a ligand which is first bound to the polypeptide.

20 Other objects of the invention are methods of the above, wherein the polypeptide is attached to a solid support.

Other objects of the invention are methods of the above, wherein the compound is attached to a solid support.

25 Another object of the invention is a method of screening for therapeutic agents useful in the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising the steps of (i) contacting a test compound with a PDE11A polynucleotide, (ii) detect binding of
30 said test compound to said PDE11A polynucleotide. Compounds that, e.g., bind to

the PDE11A polynucleotide are potential therapeutic agents for the treatment of such diseases.

5 Another object of the invention is the method of the above, wherein the nucleic acid molecule is RNA.

Another object of the invention is a method of the above, wherein the contacting step is in or at the surface of a cell.

10 Another object of the invention is a method of the above, wherein the contacting step is in a cell-free system.

Another object of the invention is a method of the above, wherein the polynucleotide is coupled to a detectable label.

15 Another object of the invention is a method of the above, wherein the test compound is coupled to a detectable label.

20 Another object of the invention is a method of diagnosing a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising the steps of (i) determining the amount of a PDE11A polynucleotide in a sample taken from said mammal, (ii) determining the amount of PDE11A polynucleotide in healthy and/or diseased mammal. A disease is
25 diagnosed, e.g., if there is a substantial similarity in the amount of PDE11A polynucleotide in said test mammal as compared to a diseased mammal.

Another object of the invention is a pharmaceutical composition for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral
30 and central nervous system, cardiovascular diseases, cancer, liver disease and genito-

urinary diseases in a mammal comprising a therapeutic agent which binds to a PDE11A polypeptide.

5 Another object of the invention is a pharmaceutical composition for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising a therapeutic agent which regulates the activity of a PDE11A polypeptide.

10 Another object of the invention is a pharmaceutical composition for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising a therapeutic agent which regulates the activity of a PDE11A polypeptide, wherein said therapeutic agent is (i) a small
15 molecule, (ii) an RNA molecule, (iii) an antisense oligonucleotide, (iv) a polypeptide, (v) an antibody, or (vi) a ribozyme.

Another object of the invention is a pharmaceutical composition for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral
20 and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising a PDE11A polynucleotide.

Another object of the invention is a pharmaceutical composition for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral
25 and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising a PDE11A polypeptide.

Another object of the invention is the use of regulators of a PDE11A for the preparation of a pharmaceutical composition for the treatment of a disease comprised
30 in a group of diseases consisting of disorders of the peripheral and central nervous

system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal.

5 Another object of the invention is a method for the preparation of a pharmaceutical composition useful for the treatment of a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal comprising the steps of (i) identifying a regulator of PDE11A, (ii) determining whether said regulator ameliorates the symptoms of a disease comprised in a group of diseases
10 consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases in a mammal; and (iii) combining of said regulator with an acceptable pharmaceutical carrier.

15 Another object of the invention is the use of a regulator of PDE11A for the regulation of PDE11A activity in a mammal having a disease comprised in a group of diseases consisting of disorders of the peripheral and central nervous system, cardiovascular diseases, cancer, liver disease and genito-urinary diseases.

20 The examples below are provided to illustrate the subject invention. These examples are provided by way of illustration and are not included for the purpose of limiting the invention.

Examples

Example 1: Search for homologous sequences in public sequence data bases

5 The degree of homology can readily be calculated by known methods. Preferred methods to determine homology are designed to give the largest match between the sequences tested. Methods to determine homology are codified in publicly available computer programs such as BestFit, BLASTP, BLASTN, and FASTA. The BLAST programs are publicly available from NCBI and other sources in the internet.

10

For PDE11A the following hits to known sequences were identified by using the BLAST algorithm [Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ; Nucleic Acids Res 1997 Sep 1; 25(17): 3389-402] and the following set of parameters: matrix = BLOSUM62 and low complexity filter. The following
15 databases were searched: NCBI (non-redundant database) and DERWENT patent database (Geneseq).

The following hits were found:

20 >NA2000:AAA49972 Aaa49972 Human cyclic nucleotide phosphodiesterase HSPDE10A1 cDNA. 10/2000, Length = 1784, Score = 3537 bits (1784), Expect = 0.0, Identities = 1784/1784 (100%)

25 >emb|AJ251509.1|HSA251509 Homo sapiens mRNA for cyclic nucleotide phosphodiesterase 11A1 (PDE11A gene), Length = 1784, Score = 3537 bits (1784), Expect = 0.0, Identities = 1784/1784 (100%)

30 >NA2001:AAH78232 Aah78232 Nucleotide sequence of a human phosphodiesterase polypeptide. 11/2001, Length = 2994, Score = 3299 bits (1664), Expect = 0.0
Identities = 1664/1664 (100%)

35 >NA2001:AAH46709 Aah46709 Human type 11 phosphodiesterase coding sequence SEQ ID NO: 3. 9/2001, Length = 3507, Score = 3299 bits (1664), Expect = 0.0

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Identities = 1664/1664 (100%)

5 >NA2001:AAH46708 Aah46708 Human type 11 phosphodiesterase coding
sequence SEQ ID NO: 1. 9/2001, Length = 4476, Score = 3299 bits
(1664), Expect = 0.0, Identities = 1664/1664 (100%)

10 >ref|NM_016953.2| Homo sapiens phosphodiesterase 11A (PDE11A), mRNA,
Length = 4476, Score = 3299 bits (1664), Expect = 0.0, Identities =
1664/1664 (100%)

>dbj|AB038041.1|AB038041 Homo sapiens HSPDE11A mRNA for
phosphodiesterase 11A2, complete cds, Length = 3507, Score = 3299
bits (1664), Expect = 0.0, Identities = 1664/1664 (100%)

15 >dbj|AB036704.1|AB036704 Homo sapiens HSPDE11A mRNA for
phosphodiesterase 11A, complete cds, Length = 4476, Score = 3299
bits (1664), Expect = 0.0, Identities = 1664/1664 (100%)

20 >NA2001:AAH78223 Aah78223 Nucleotide sequence of a human
phosphodiesterase polypeptide. 11/2001, Length = 2513, Score = 3128
bits (1578), Expect = 0.0, Identities = 1584/1586 (99%)

25 >gb|AF281865.1|AF281865 Homo sapiens cAMP/cGMP phosphodiesterase
11A2 mRNA, complete cds, alternatively spliced, Length = 2141, Score
= 2997 bits (1512), Expect = 0.0, Identities = 1515/1516 (99%)

30 >emb|AJ278682.1|HSA278682 Homo sapiens mRNA for cAMP/cGMP cyclic
nucleotide phosphodiesterase, 11A3 (PDE11A3 gene), Length = 2502,
Score = 2997 bits (1512), Expect = 0.0, Identities = 1515/1516 (99%)

Example 2: Expression profiling

35 Total cellular RNA was isolated from cells by one of two standard methods: 1)
guanidine isothiocyanate/Cesium chloride density gradient centrifugation [Kellogg,
(1990)] ; or with the Tri-Reagent protocol according to the manufacturer's
specifications (Molecular Research Center, Inc., Cincinnati, Ohio). Total RNA
prepared by the Tri-reagent protocol was treated with DNase I to remove genomic
DNA contamination.

For relative quantitation of the mRNA distribution of PDE11A, total RNA from each cell or tissue source was first reverse transcribed. 85 µg of total RNA was reverse transcribed using 1 µmole random hexamer primers, 0.5 mM each of dATP, dCTP, dGTP and dTTP (Qiagen, Hilden, Germany), 3000 U RnaseQut (Invitrogen, Groningen, Netherlands) in a final volume of 680 µl. The first strand synthesis buffer and Omniscript reverse transcriptase (2 u/µl) were from (Qiagen, Hilden, Germany). The reaction was incubated at 37°C for 90 minutes and cooled on ice. The volume was adjusted to 6800 µl with water, yielding a final concentration of 12.5 ng/µl of starting RNA.

For relative quantitation of the distribution of PDE11A mRNA in cells and tissues the Perkin Elmer ABI Prism RTM. 7700 Sequence Detection system or Biorad iCycler was used according to the manufacturer's specifications and protocols. PCR reactions were set up to quantitate PDE11A and the housekeeping genes HPRT (hypoxanthine phosphoribosyltransferase), GAPDH (glyceraldehyde-3-phosphate dehydrogenase), β-actin, and others. Forward and reverse primers and probes for PDE11A were designed using the Perkin Elmer ABI Primer Express™ software and were synthesized by TibMolBiol (Berlin, Germany). The PDE11A forward primer sequence was: Primer1 (SEQ ID NO: 3). The PDE11A reverse primer sequence was Primer2 (SEQ ID NO: 5). Probe1 (SEQ ID NO: 4), labelled with FAM (carboxyfluorescein succinimidyl ester) as the reporter dye and TAMRA (carboxytetramethylrhodamine) as the quencher, is used as a probe for PDE11A. The following reagents were prepared in a total of 25 µl : 1x TaqMan buffer A, 5.5 mM MgCl₂, 200 nM of dATP, dCTP, dGTP, and dUTP, 0.025 U/µl AmpliTaq Gold™, 0.01 U/ µl AmpErase and Probe1 (SEQ ID NO: 4), PDE11A forward and reverse primers each at 200 nM, 200 nM PDE11A FAM/TAMRA-labelled probe, and 5 µl of template cDNA. Thermal cycling parameters were 2 min at 50°C, followed by 10 min at 95°C, followed by 40 cycles of melting at 95°C for 15 sec and annealing/extending at 60°C for 1 min.

Calculation of corrected CT values

The CT (threshold cycle) value is calculated as described in the "Quantitative determination of nucleic acids" section. The CF-value (factor for threshold cycle correction) is calculated as follows :

1. PCR reactions were set up to quantitate the housekeeping genes (HKG) for each cDNA sample.
2. CT_{HKG} -values (threshold cycle for housekeeping gene) were calculated as described in the "Quantitative determination of nucleic acids" section.
3. CT_{HKG} -mean values (CT mean value of all HKG tested on one cDNAs) of all HKG for each cDNA are calculated (n = number of HKG):

$$CT_{HKG-n}\text{-mean value} = (CT_{HKG1}\text{-value} + CT_{HKG2}\text{-value} + \dots + CT_{HKG-n}\text{-value}) / n$$

4. CT_{panel} mean value (CT mean value of all HKG in all tested cDNAs) =

$$(CT_{HKG1}\text{-mean value} + CT_{HKG2}\text{-mean value} + \dots + CT_{HKG-y}\text{-mean value}) / y$$

(y = number of cDNAs)

5. CF_{cDNA-n} (correction factor for cDNA n) = $CT_{\text{panel}}\text{-mean value} - CT_{HKG-n}\text{-mean value}$

6. CT_{cDNA-n} (CT value of the tested gene for the cDNA n) + CF_{cDNA-n} (correction factor for cDNA n) = $CT_{\text{cor-cDNA-n}}$ (corrected CT value for a gene on cDNA n)

Calculation of relative expression

Definition : highest $CT_{\text{cor-cDNA-n}} \neq 40$ is defined as $CT_{\text{cor-cDNA}} [\text{high}]$

Relative Expression = $2^{(CT_{\text{cor-cDNA}} [\text{high}] - CT_{\text{cor-cDNA-n}})}$

5

Tissues

The expression of PDE11A was investigated in the following tissues: breast, testis, prostate, liver, skeletal muscle, fetal kidney, thyroid, fetal brain, salivary gland, mammary gland, heart ventricle (left), HEK 293 cells, pericardium, adipose, HEP G2 cells, Alzheimer cerebral cortex, trachea, thalamus, small intestine, kidney, adrenal gland, colon, interventricular septum, MDA MB 231 cells (breast tumor), heart atrium (right), fetal liver, heart atrium (left), uterus, occipital lobe, precentral gyrus, temporal lobe, heart, thyroid tumor, postcentral gyrus, hippocampus, pons, stomach, cerebral meninges, cerebral cortex, liver liver cirrhosis, parietal lobe, esophagus, frontal lobe, pancreas, lung tumor, brain, cervix, HUVEC cells, breast tumor, thymus, spinal cord, cerebral peduncles, Alzheimer brain frontal lobe, vermis cerebelli, colon tumor, lung, fetal lung, bladder, cerebellum (left), spleen, fetal heart, tonsilla cerebelli, bone marrow, corpus callosum, coronary artery smooth muscle primary cells, pancreas liver cirrhosis, Jurkat (T-cells), HeLa cells (cervix tumor), Alzheimer brain, cerebellum (right), placenta, leukocytes (peripheral blood), cerebellum, spleen liver cirrhosis

10

15

20

Expression profile

25

The results of the the mRNA-quantification (expression profiling) is shown in Table 1.

Table 1: Relative expression of PDE11A in various human tissues.

	breast	1380
	testis	1278
5	prostate	1105
	liver	873
	skeletal muscle	820
	fetal kidney	724
	thyroid	205
10	fetal brain	194
	salivary gland	160
	mammary gland	128
	heart ventricle (left)	119
	HEK 293 cells	119
15	pericardium	111
	adipose	107
	HEP G2 cells	103
	Alzheimer cerebral cortex	95
	trachea	87
20	thalamus	79
	small intestine	61
	kidney	53
	adrenal gland	51
	colon	49
25	interventricular septum	46
	MDA MB 231 cells (breast tumor)	45
	heart atrium (right)	44
	fetal liver	42
	heart atrium (left)	38
30	uterus	30
	occipital lobe	29
	precentral gyrus	29
	temporal lobe	28
	heart	27
35	thyroid tumor	27
	postcentral gyrus	26
	hippocampus	26
	pons	26
	stomach	24
40	cerebral meninges	23
	cerebral cortex	21
	liver liver cirrhosis	18
	parietal lobe	18
	esophagus	16
45	frontal lobe	14
	pancreas	14
	lung tumor	13
	brain	12
	cervix	12
50	HUVEC cells	11

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	breast tumor	11
	thymus	9
	spinal cord	8
	cerebral peduncles	8
5	Alzheimer brain frontal lobe	8
	vermis cerebelli	7
	colon tumor	6
	lung	0
	fetal lung	5
10	bladder	5
	cerebellum (left)	4
	spleen	4
	fetal heart	3
	tonsilla cerebelli	3
15	bone marrow	3
	corpus callosum	2
	coronary artery smooth muscle	
	primary cells	0
	pancreas liver cirrhosis	2
20	Jurkat (T-cells)	1
	HeLa cells (cervix tumor)	1
	Alzheimer brain	1
	cerebellum (right)	1
	placenta	0
25	leukocytes (peripheral blood)	0
	cerebellum	0
	spleen liver cirrhosis	0

Example 3: Antisense Analysis

30

Knowledge of the correct, complete cDNA sequence coding for PDE11A enables its use as a tool for antisense technology in the investigation of gene function. Oligonucleotides, cDNA or genomic fragments comprising the antisense strand of a polynucleotide coding for PDE11A are used either in vitro or in vivo to inhibit

35 translation of the mRNA. Such technology is now well known in the art, and antisense molecules can be designed at various locations along the nucleotide sequences. By treatment of cells or whole test animals with such antisense sequences, the gene of interest is effectively turned off. Frequently, the function of the gene is ascertained by observing behavior at the intracellular, cellular, tissue or

40 organismal level (e.g., lethality, loss of differentiated function, changes in morphology, etc.).

In addition to using sequences constructed to interrupt transcription of a particular open reading frame, modifications of gene expression is obtained by designing antisense sequences to intron regions, promoter/enhancer elements, or even to trans-
5 acting regulatory genes.

Example 4: Expression of PDE11A

Expression of PDE11A is accomplished by subcloning the cDNAs into appropriate
10 expression vectors and transfecting the vectors into expression hosts such as, e.g., *E. coli*. In a particular case, the vector is engineered such that it contains a promoter for β -galactosidase, upstream of the cloning site, followed by sequence containing the amino-terminal Methionine and the subsequent seven residues of β -galactosidase. Immediately following these eight residues is an engineered bacteriophage promoter
15 useful for artificial priming and transcription and for providing a number of unique endonuclease restriction sites for cloning.

Induction of the isolated, transfected bacterial strain with Isopropyl- β -D-thio-
20 galactopyranoside (IPTG) using standard methods produces a fusion protein corresponding to the first seven residues of β -galactosidase, about 15 residues of "linker", and the peptide encoded within the cDNA. Since cDNA clone inserts are generated by an essentially random process, there is probability of 33% that the included cDNA will lie in the correct reading frame for proper translation. If the cDNA is not in the proper reading frame, it is obtained by deletion or insertion of the
25 appropriate number of bases using well known methods including in vitro mutagenesis, digestion with exonuclease III or mung bean nuclease, or the inclusion of an oligonucleotide linker of appropriate length.

The PDE11A cDNA is shuttled into other vectors known to be useful for expression
30 of proteins in specific hosts. Oligonucleotide primers containing cloning sites as well as a segment of DNA (about 25 bases) sufficient to hybridize to stretches at both

ends of the target cDNA is synthesized chemically by standard methods. These primers are then used to amplify the desired gene segment by PCR. The resulting gene segment is digested with appropriate restriction enzymes under standard conditions and isolated by gel electrophoresis. Alternately, similar gene segments
5 are produced by digestion of the cDNA with appropriate restriction enzymes. Using appropriate primers, segments of coding sequence from more than one gene are ligated together and cloned in appropriate vectors. It is possible to optimize expression by construction of such chimeric sequences.

10 Suitable expression hosts for such chimeric molecules include, but are not limited to, mammalian cells such as Chinese Hamster Ovary (CHO) and human 293 cells., insect cells such as Sf9 cells, yeast cells such as *Saccharomyces cerevisiae* and bacterial cells such as *E. coli*. For each of these cell systems, a useful expression vector also includes an origin of replication to allow propagation in bacteria, and a
15 selectable marker such as the β -lactamase antibiotic resistance gene to allow plasmid selection in bacteria. In addition, the vector may include a second selectable marker such as the neomycin phosphotransferase gene to allow selection in transfected eukaryotic host cells. Vectors for use in eukaryotic expression hosts require RNA processing elements such as 3' polyadenylation sequences if such are not part of the
20 cDNA of interest.

Additionally, the vector contains promoters or enhancers which increase gene expression. Such promoters are host specific and include MMTV, SV40, and metallothionine promoters for CHO cells; trp, lac, tac and T7 promoters for bacterial
25 hosts; and alpha factor, alcohol oxidase and PGH promoters for yeast. Transcription enhancers, such as the rous sarcoma virus enhancer, are used in mammalian host cells. Once homogeneous cultures of recombinant cells are obtained through standard culture methods, large quantities of recombinantly produced PDE11A are recovered from the conditioned medium and analyzed using chromatographic
30 methods known in the art. For example, PDE11A can be cloned into the expression vector pcDNA3, as exemplified herein. This product can be used to transform, for

example, HEK293 or COS by methodology standard in the art. Specifically, for example, using Lipofectamine (Gibco BRL catalog no. 18324-020) mediated gene transfer.

5 Example 5: Isolation of Recombinant PDE11A

PDE11A is expressed as a chimeric protein with one or more additional polypeptide domains added to facilitate protein purification. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals [Appa Rao, 1997] and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Washington). The inclusion of a cleavable linker sequence such as Factor Xa or enterokinase (Invitrogen, Groningen, The Netherlands) between the purification domain and the PDE11A sequence is useful to facilitate expression of PDE11A.

The following example provides a method for purifying PDE11A.

PDE11A is generated using the baculovirus expression system BAC-TO-BAC (GIBCO BRL) based on *Autographa californica* nuclear polyhedrosis virus (AcNPV) infection of *Spodoptera frugiperda* insect cells (Sf9 cells).

cDNA encoding PDE is cloned into either the donor plasmid pFASTBAC1 or pFASTBAC-HT which contain a mini-Tn7 transposition element. The recombinant plasmid is transformed into DH10BAC competent cells which contain the parent bacmid bMON14272 (AcNPV infectious DNA) and a helper plasmid. The mini-Tn7 element on the pFASTBAC donor can transpose to the attTn7 attachment site on the bacmid thus introducing the PDE gene into the viral genome. Colonies containing recombinant bacmids are identified by disruption of the *lacZ* gene. The PDE/bacmid construct can then be isolated and infected into insect cells (Sf9 cells) resulting in the production of infectious recombinant baculovirus particles and expression of either

unfused recombinant enzyme (pFastbac1) or PDE11A-His fusion protein (pFastbacHT).

Cells are harvested and extracts prepared 24, 48 and 72 hours after transfection. Expression of PDE11A is confirmed by coomassie staining after sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and western blotting onto a PVDF membrane of an unstained SDS-PAGE. The PDE-His fusion protein is detected due to the interaction between the Ni-NTA HRP conjugate and the His-tag which is fused to PDE11A.

Example 6: Production of PDE11A Specific Antibodies

Two approaches are utilized to raise antibodies to PDE11A, and each approach is useful for generating either polyclonal or monoclonal antibodies. In one approach, denatured protein from reverse phase HPLC separation is obtained in quantities up to 75 mg. This denatured protein is used to immunize mice or rabbits using standard protocols; about 100 µg are adequate for immunization of a mouse, while up to 1 mg might be used to immunize a rabbit. For identifying mouse hybridomas, the denatured protein is radioiodinated and used to screen potential murine B-cell hybridomas for those which produce antibody. This procedure requires only small quantities of protein, such that 20 mg is sufficient for labeling and screening of several thousand clones.

In the second approach, the amino acid sequence of an appropriate PDE11A domain, as deduced from translation of the cDNA, is analyzed to determine regions of high antigenicity. Oligopeptides comprising appropriate hydrophilic regions are synthesized and used in suitable immunization protocols to raise antibodies. The optimal amino acid sequences for immunization are usually at the C-terminus, the N-terminus and those intervening, hydrophilic regions of the polypeptide which are likely to be exposed to the external environment when the protein is in its natural conformation.

Typically, selected peptides, about 15 residues in length, are synthesized using an Applied Biosystems Peptide Synthesizer Model 431A using fmoc-chemistry and coupled to keyhole limpet hemocyanin (KLH; Sigma, St. Louis, MO) by reaction with M-maleimidobenzoyl-N-hydroxysuccinimide ester, MBS. If necessary, a cysteine is introduced at the N-terminus of the peptide to permit coupling to KLH. Rabbits are immunized with the peptide-KLH complex in complete Freund's adjuvant. The resulting antisera are tested for antipeptide activity by binding the peptide to plastic, blocking with 1% bovine serum albumin, reacting with antisera, washing and reacting with labeled (radioactive or fluorescent), affinity purified, specific goat anti-rabbit IgG.

Hybridomas are prepared and screened using standard techniques. Hybridomas of interest are detected by screening with labeled PDE11A to identify those fusions producing the monoclonal antibody with the desired specificity. In a typical protocol, wells of plates (FAST; Becton-Dickinson, Palo Alto, CA) are coated during incubation with affinity purified, specific rabbit anti-mouse (or suitable antispecies 1 g) antibodies at 10 mg/ml. The coated wells are blocked with 1% bovine serum albumin, (BSA), washed and incubated with supernatants from hybridomas. After washing the wells are incubated with labeled PDE11A at 1 mg/ml. Supernatants with specific antibodies bind more labeled PDE11A than is detectable in the background. Then clones producing specific antibodies are expanded and subjected to two cycles of cloning at limiting dilution. Cloned hybridomas are injected into pristane-treated mice to produce ascites, and monoclonal antibody is purified from mouse ascitic fluid by affinity chromatography on Protein A. Monoclonal antibodies with affinities of at least

10^8 M^{-1} , preferably 10^9 to 10^{10} M^{-1} or stronger, are typically made by standard procedures.

Example 7: Diagnostic Test Using PDE11A Specific Antibodies

Particular PDE11A antibodies are useful for investigating signal transduction and the diagnosis of infectious or hereditary conditions which are characterized by differences in the amount or distribution of PDE11A or downstream products of an active signaling cascade.

Diagnostic tests for PDE11A include methods utilizing antibody and a label to detect PDE11A in human body fluids, membranes, cells, tissues or extracts of such. The polypeptides and antibodies of the present invention are used with or without modification. Frequently, the polypeptides and antibodies are labeled by joining them, either covalently or noncovalently, with a substance which provides for a detectable signal. A wide variety of labels and conjugation techniques are known and have been reported extensively in both the scientific and patent literature. Suitable labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent agents, chemiluminescent agents, chromogenic agents, magnetic particles and the like.

A variety of protocols for measuring soluble or membrane-bound PDE11A, using either polyclonal or monoclonal antibodies specific for the protein, are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA) and fluorescent activated cell sorting (FACS). A two-site monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on PDE11A is preferred, but a competitive binding assay may be employed.

Example 8: Purification of Native PDE11A Using Specific Antibodies

Native or recombinant PDE11A is purified by immunoaffinity chromatography using antibodies specific for PDE11A. In general, an immunoaffinity column is

constructed by covalently coupling the anti-TRH antibody to an activated chromatographic resin.

5 Polyclonal immunoglobulins are prepared from immune sera either by precipitation with ammonium sulfate or by purification on immobilized Protein A (Pharmacia LKB Biotechnology, Piscataway N.J.). Likewise, monoclonal antibodies are prepared from mouse ascites fluid by ammonium sulfate precipitation or chromatography on immobilized Protein A. Partially purified immunoglobulin is covalently attached to a chromatographic resin such as CnBr-activated Sepharose
10 (Pharmacia LKB Biotechnology). The antibody is coupled to the resin, the resin is blocked, and the derivative resin is washed according to the manufacturer's instructions.

15 Such immunoaffinity columns are utilized in the purification of PDE11A by preparing a fraction from cells containing PDE11A in a soluble form. This preparation is derived by solubilization of whole cells or of a subcellular fraction obtained via differential centrifugation (with or without addition of detergent) or by other methods well known in the art. Alternatively, soluble PDE11A containing a signal sequence is secreted in useful quantity into the medium in which the cells are grown.

20 A soluble PDE11A-containing preparation is passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of PDE11A (e.g., high ionic strength buffers in the presence of detergent). Then, the column is eluted under conditions that disrupt antibody/protein
25 binding (e.g., a buffer of pH 2-3 or a high concentration of a chaotrope such as urea or thiocyanate ion), and PDE11A is collected.

Example 9: Drug Screening

30 This invention is particularly useful for screening therapeutic compounds by using PDE11A or fragments thereof in any of a variety of drug screening techniques.

The following example provides a system for drug screening measuring the phosphodiesterase activity.

5 The phosphodiesterase activity of the crude extracts is measured and confirmed that the PDE cDNA encodes a phosphodiesterase which is able to hydrolyze cAMP or cGMP or both.

10 The recombinant PDE-His fusion protein can be purified from the crude lysate by metal-affinity chromatography using Ni-NTA agarose. This allows the specific retention of the recombinant material (since this is fused to the His-tag) whilst the endogenous insect proteins are washed off. The recombinant material is then eluted by competition with imidazol.

15 The phosphodiesterase activity of the recombinant protein is assayed using a commercially available SPA (scintillation proximity assay) kit (Amersham Pharmacia). The PDE enzyme hydrolyzes cyclic nucleotides, e.g. cAMP and cGMP to their linear counterparts. The SPA assay utilizes the tritiated cyclic nucleotides [^3H]cAMP or [^3H]cGMP, and is based upon the selective interaction of the tritiated non cyclic product with the SPA beads whereas the cyclic substrates are not
20 effectively binding. Radiolabelled product bound to the scintillation beads generates light that can be analyzed in a scintillation counter.

Example 10: Rational Drug Design

25 The goal of rational drug design is to produce structural analogs of biologically active polypeptides of interest or of small molecules with which they interact, agonists, antagonists, or inhibitors. Any of these examples are used to fashion drugs which are more active or stable forms of the polypeptide or which enhance or
30 interfere with the function of a polypeptide in vivo.

In one approach, the three-dimensional structure of a protein of interest, or of a protein-inhibitor complex, is determined by x-ray crystallography, by computer modeling or, most typically, by a combination of the two approaches. Both the shape and charges of the polypeptide must be ascertained to elucidate the structure and to determine active site(s) of the molecule. Less often, useful information regarding the structure of a polypeptide is gained by modeling based on the structure of homologous proteins. In both cases, relevant structural information is used to design efficient inhibitors. Useful examples of rational drug design include molecules which have improved activity or stability or which act as inhibitors, agonists, or antagonists of native peptides.

It is also possible to isolate a target-specific antibody, selected by functional assay, as described above, and then to solve its crystal structure. This approach, in principle, yields a pharmacore upon which subsequent drug design is based. It is possible to bypass protein crystallography altogether by generating anti-idiotypic antibodies (anti-ids) to a functional, pharmacologically active antibody. As a mirror image of a mirror image, the binding site of the anti-ids is expected to be an analog of the original receptor. The anti-id is then used to identify and isolate peptides from banks of chemically or biologically produced peptides. The isolated peptides then act as the pharmacore.

By virtue of the present invention, sufficient amount of polypeptide are made available to perform such analytical studies as X-ray crystallography. In addition, knowledge of the PDE11A amino acid sequence provided herein provides guidance to those employing computer modeling techniques in place of or in addition to x-ray crystallography.

Example 11: Identification of Other Members of the Signal Transduction Complex

Labeled PDE11A is useful as a reagent for the purification of molecules with which it interacts. In one embodiment of affinity purification, PDE11A is covalently coupled to a chromatography column. Cell-free extract derived from synovial cells or putative target cells is passed over the column, and molecules with appropriate affinity bind to PDE11A. PDE11A-complex is recovered from the column, and the PDE11A-binding ligand disassociated and subjected to N-terminal protein sequencing. The amino acid sequence information is then used to identify the captured molecule or to design degenerate oligonucleotide probes for cloning the relevant gene from an appropriate cDNA library.

In an alternate method, antibodies are raised against PDE11A, specifically monoclonal antibodies. The monoclonal antibodies are screened to identify those which inhibit the binding of labeled PDE11A. These monoclonal antibodies are then used therapeutically.

Example 12: Use and Administration of Antibodies, Inhibitors, or Antagonists

Antibodies, inhibitors, or antagonists of PDE11A or other treatments and compounds that are limiters of signal transduction (LSTs), provide different effects when administered therapeutically. LSTs are formulated in a nontoxic, inert, pharmaceutically acceptable aqueous carrier medium preferably at a pH of about 5 to 8, more preferably 6 to 8, although pH may vary according to the characteristics of the antibody, inhibitor, or antagonist being formulated and the condition to be treated. Characteristics of LSTs include solubility of the molecule, its half-life and antigenicity/immunogenicity. These and other characteristics aid in defining an effective carrier. Native human proteins are preferred as LSTs, but organic or synthetic molecules resulting from drug screens are equally effective in particular situations.

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LSTs are delivered by known routes of administration including but not limited to topical creams and gels; transmucosal spray and aerosol; transdermal patch and bandage; injectable, intravenous and lavage formulations; and orally administered liquids and pills particularly formulated to resist stomach acid and enzymes. The particular formulation, exact dosage, and route of administration is determined by the attending physician and varies according to each specific situation.

Such determinations are made by considering multiple variables such as the condition to be treated, the LST to be administered, and the pharmacokinetic profile of a particular LST. Additional factors which are taken into account include severity of the disease state, patient's age, weight, gender and diet, time and frequency of LST administration, possible combination with other drugs, reaction sensitivities, and tolerance/response to therapy. Long acting LST formulations might be administered every 3 to 4 days, every week, or once every two weeks depending on half-life and clearance rate of the particular LST.

Normal dosage amounts vary from 0.1 to 10^5 μ g, up to a total dose of about 1 g, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature; see U.S. Pat. Nos. 4,657,760; 5,206,344; or 5,225,212. Those skilled in the art employ different formulations for different LSTs. Administration to cells such as nerve cells necessitates delivery in a manner different from that to other cells such as vascular endothelial cells.

It is contemplated that abnormal signal transduction, trauma, or diseases which trigger PDE11A activity are treatable with LSTs. These conditions or diseases are specifically diagnosed by the tests discussed above, and such testing should be performed in suspected cases of viral, bacterial or fungal infections, allergic responses, mechanical injury associated with trauma, hereditary diseases, lymphoma or carcinoma, or other conditions which activate the genes of lymphoid or neuronal tissues.

Example 13: Production of Non-human Transgenic Animals

Animal model systems which elucidate the physiological and behavioral roles of the PDE11A are produced by creating nonhuman transgenic animals in which the activity of the PDE11A is either increased or decreased, or the amino acid sequence of the expressed PDE11A is altered, by a variety of techniques. Examples of these techniques include, but are not limited to: 1) Insertion of normal or mutant versions of DNA encoding a PDE11A, by microinjection, electroporation, retroviral transfection or other means well known to those skilled in the art, into appropriately fertilized embryos in order to produce a transgenic animal or 2) homologous recombination of mutant or normal, human or animal versions of these genes with the native gene locus in transgenic animals to alter the regulation of expression or the structure of these PDE11A sequences. The technique of homologous recombination is well known in the art. It replaces the native gene with the inserted gene and hence is useful for producing an animal that cannot express native PDE11As but does express, for example, an inserted mutant PDE11A, which has replaced the native PDE11A in the animal's genome by recombination, resulting in underexpression of the transporter. Microinjection adds genes to the genome, but does not remove them, and the technique is useful for producing an animal which expresses its own and added PDE11A, resulting in overexpression of the PDE11A.

One means available for producing a transgenic animal, with a mouse as an example, is as follows: Female mice are mated, and the resulting fertilized eggs are dissected out of their oviducts. The eggs are stored in an appropriate medium such as cesiumchloride M2 medium. DNA or cDNA encoding PDE11A is purified from a vector by methods well known to the one skilled in the art. Inducible promoters may be fused with the coding region of the DNA to provide an experimental means to regulate expression of the transgene. Alternatively or in addition, tissue specific regulatory elements may be fused with the coding region to permit tissue-specific expression of the transgene. The DNA, in an appropriately buffered solution, is put into a microinjection needle (which may be made from capillary tubing using a piper

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puller) and the egg to be injected is put in a depression slide. The needle is inserted into the pronucleus of the egg, and the DNA solution is injected. The injected egg is then transferred into the oviduct of a pseudopregnant mouse which is a mouse stimulated by the appropriate hormones in order to maintain false pregnancy, where
5 it proceeds to the uterus, implants, and develops to term. As noted above, microinjection is not the only method for inserting DNA into the egg but is used here only for exemplary purposes.

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